

Modeling and simulation of gene regulatory networks 1

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INRIA Grenoble - Rhône-Alpes and IBIS



- IBIS: systems biology group at INRIA/Université Joseph Fourier/CNRS
 - Analysis of bacterial regulatory networks by means of models and experiments
 - Biologists, computer scientists, mathematicians, physicists, ...

http://ibis.inrialpes.fr







Overview

- 1. Gene regulatory networks in bacteria
- 2. Deterministic modeling of gene regulatory networks
- 3. Qualitative modeling of gene regulatory networks
- 4. Stochastic modeling of gene regulatory networks
- 5. Some current issues and perspectives



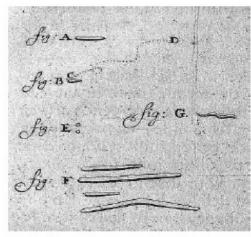


Bacteria

 Bacteria were first observed by Antonie van Leeuwenhoek, using a single-lens microscope of his own design



http://commons.wikimedia.org/



van Leeuwenhoek A (1684), Philosophical Transactions (1683–1775) 14: 568–574

www.euronet.nl/users/warnar/leeuwenhoek.html.

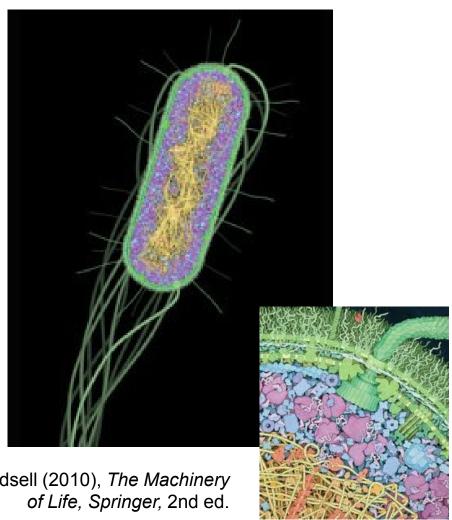
"In the morning I used to rub my teeth with salt and rinse my mouth with water and after eating to clean my molars with a toothpick.... I then most always saw, with great wonder, that in the said matter there were many very **little living animalcules**, very prettily amoving. The biggest sort had a very strong and swift motion, and shot through the water like a pike does through the water; mostly these were of small numbers."

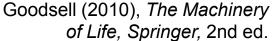




Bacteria are complex living systems

- Bacterial cells are complex biochemical and biophysical machines
 - Wide range of shapes, typically 0.5-5 µm in length
 - 10⁶ bacterial cells in 1 ml of fresh water
 - 10 times as much bacterial cells as human cells in human body



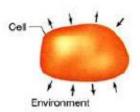






Bacteria are complex living systems

- Bacterial cells are complex biochemical and biophysical machines
- Bacteria possess
 characteristics shared by most living systems:
 - Metabolism
 - Growth and reproduction
 - Differentiation
 - Communication
 - Evolution



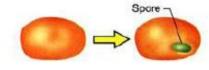
1. Metabolism

Uptake of chemicals from the environment, their transformation within the cell, and elimination of wastes into the environment. The cell is thus an open system.



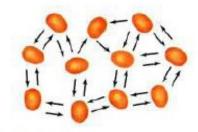
2. Reproduction (growth)

Chemicals from the environment are turned into new cells under the direction of preexisting cells.



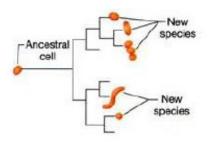
3. Differentiation

Formation of a new cell structure such as a spore, usually as part of a cellular life cycle.



4. Communication

Cells communicate or interact primarily by means of chemicals that are released or taken up.



5. Evolution

Cells evolve to display new biological properties. Phylogenetic trees show the evolutionary relationships between cells.

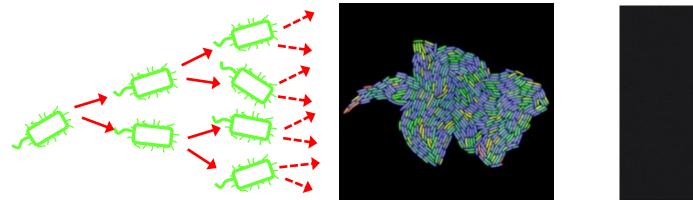
Madigan et al. (2003), Brock Biology of Microorganisms, Prentice Hall, 10th ed.





 Bacteria are unicellular organisms geared towards growth and division

Escherichia coli cells have doubling times up to 20 min





Stewart et al. (2005), PLoS Biol., 3(2): e45

 Metabolism fuels growth by production of energy and building blocks for macromolecules, using nutriments from environment

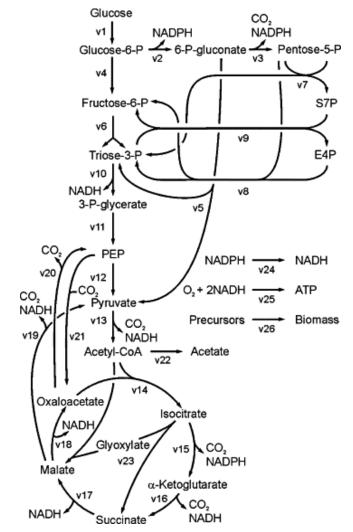
ATP, amino acids, nucleotides, ...





 Central carbon metabolism breaks down carbon sources for energy production and macromolecular synthesis

Glucose, acetate, lactose, ...



Fischer et al. (2004), Anal. Biochem., 325(2):308-16



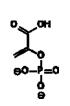


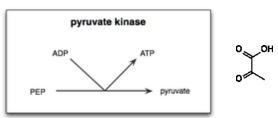
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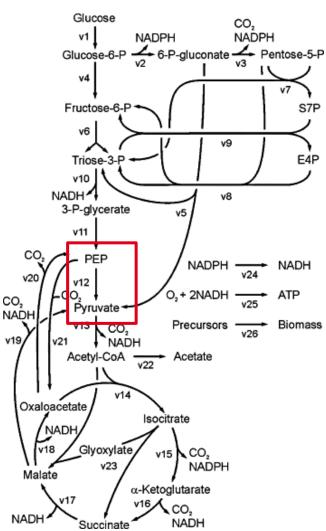
Glucose, acetate, lactose, ...

Enzymes catalyse individual steps in metabolic network

Pyruvate kinase transforms phosphoenolpyruvate (PEP) into pyruvate







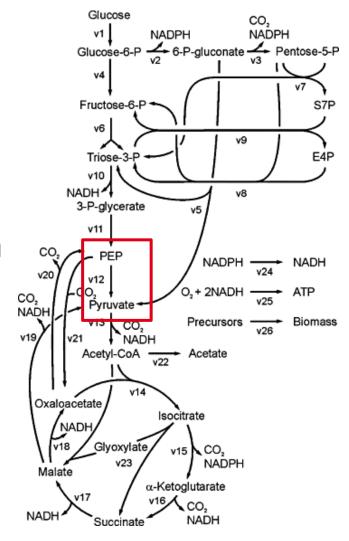




 Central carbon metabolism breaks down carbon sources for energy production and macromolecular synthesis

Glucose, acetate, lactose, ...

- Enzymes produced from information encoded in genes
 - pykF is gene encoding pyruvate kinase





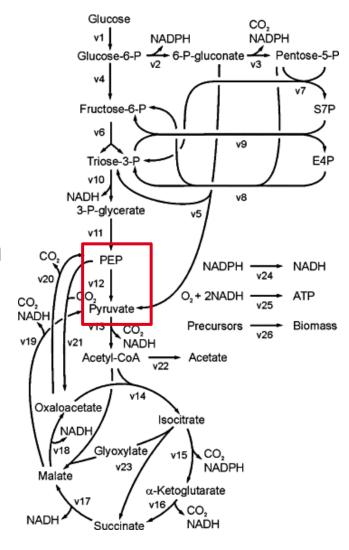


 Central carbon metabolism breaks down carbon sources for energy production and macromolecular synthesis

Glucose, acetate, lactose, ...

- Enzymes produced from information encoded in genes
 - pykF is gene encoding pyruvate kinase
 - Expression of pykF regulated by transcription factor Cra



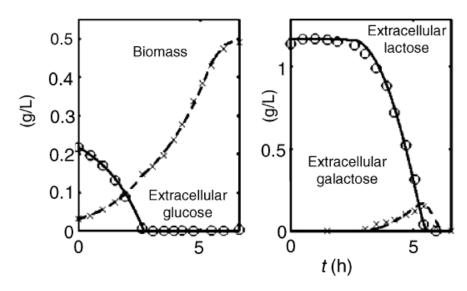






 Bacterial metabolism is flexible, allowing cells to grow on different carbon sources

Preferential utilisation: diauxic growth on glucose and lactose



Bettenbrock et al. (2006), J. Biol. Chem., 281(5):2578-84

Adaptation of bacterial physiology to different carbon sources

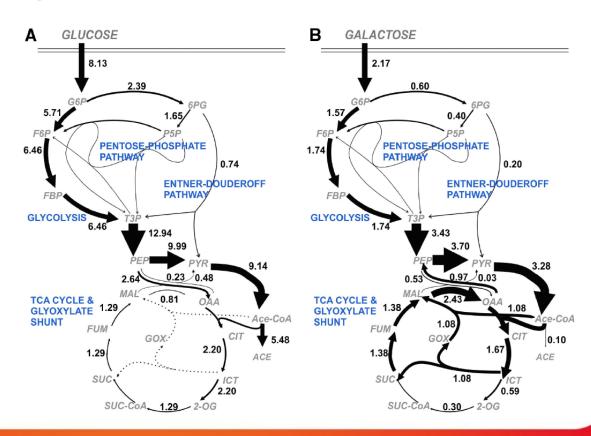




Growth transition and metabolism

Adaptation to different carbon source involves changes in metabolic fluxes

Different flux distribution in central metabolism of *E. coli* during growth on glucose and galactose



Haverkorn van Rijsewijk *et al.* (2011), *Mol. Syst. Biol.*, 7:477





Growth transition and metabolism

 Adaptation to different carbon source involves adjustment of metabolite concentrations

Different metabolite concentrations in *E. coli* cells growing on glucose and acetate

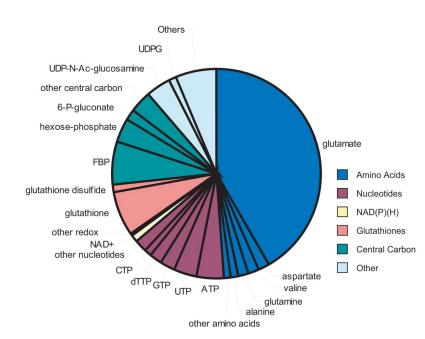


Table 1 Intracellular metabolite concentrations in glucose-fed, exponentially growing E. coli

| Metabolite | mol I⁻¹ | Metabolite | mol I⁻¹ |
|---------------------------------------|------------------------|--------------------------------|----------------------|
| Glutamate | 9.6 × 10 ⁻² | UDP-glucuronate (51) | 5.7×10^{-4} |
| Glutathione | 1.7×10^{-2} | ADP | 5.6×10^{-4} |
| Fructose-1,6-bisphosphate | 1.5×10^{-2} | Asparagine (52) | 5.1×10^{-4} |
| ATP | 9.6×10^{-3} | α-Ketoglutarate | 4.4×10^{-4} |
| UDP- <i>N</i> -acetylglucosamine (29) | 9.2×10^{-3} | Lysine (53) | 4.1×10^{-4} |
| Hexose-Pa | 8.8×10^{-3} | Proline (54) | 3.9×10^{-4} |
| UTP (30) | 8.3×10^{-3} | dTDP (55) | 3.8×10^{-4} |
| GTP (31) | 4.9×10^{-3} | Dihydroxyacetone phosphate | 3.7×10^{-4} |
| dTTP | 4.6×10^{-3} | Homocysteine (56) | 3.7×10^{-4} |
| Aspartate | 4.2×10^{-3} | CMP (57) | 3.6×10^{-4} |
| Valine (32) | 4.0×10^{-3} | Deoxyribose-5-P (58) | 3.0×10^{-4} |
| Glutamine | 3.8×10^{-3} | Isoleucine (59) + leucine (60) | 3.0×10^{-4} |
| 6-Phosphogluconate | 3.8×10^{-3} | AMP | 2.8×10^{-4} |

Bennett et al. (2009), Nat. Chem. Biol., 5(8):593-9





Growth transition and gene expression

 Adaptation to different carbon source involves adjustment of expression of enzymatic genes

Difference in expression levels of genes encoding enzymes in central metabolism of *E. coli* during growth on glucose and acetate

glk (1.1) ptsHI-crr (0.47 - 0.54)6PGnt pfkA (0.59) pfkB (1.2) tktA (1.3) tktB (1.3) tpiA (0.95) gapA (0.45) eno (0.54) (0.28)(0.29 - 0.44)pltA (4.9 acnB (6.9) icdA (1.8) fumC (2.1) sucCD (2.8-3.1)

Oh et al. (2002), J. Biol. Chem., 277(15):13175-83

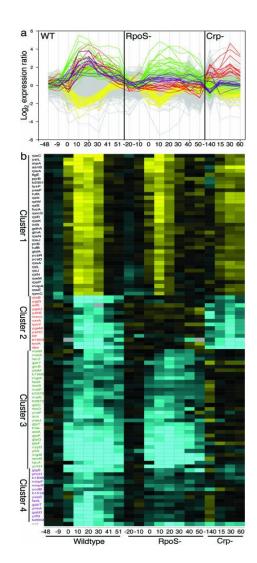




Growth transition and gene expression

 Adaptation to different carbon source involves genome-wide reorganisation of gene expression

Gene expression during glucose-lactose shift in *E. coli*



Traxler et al. (2006), Proc. Natl. Acad. Sci. USA, 103(7):2374-9

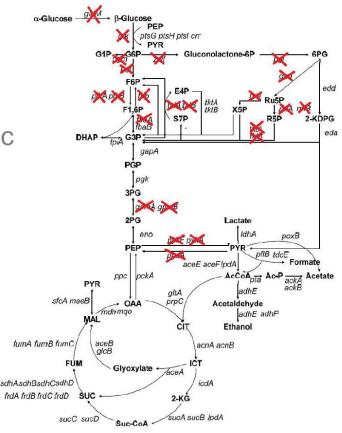




Adaptation on multiple levels

- Adaptation to different carbon source involves adjustments on multiple levels at the same time!
 - Parallel measurement of enzyme and metabolite concentrations, and metabolic fluxes in a variety of experimental conditions

Ishii et al. (2007), Science, 316(5284):593-7



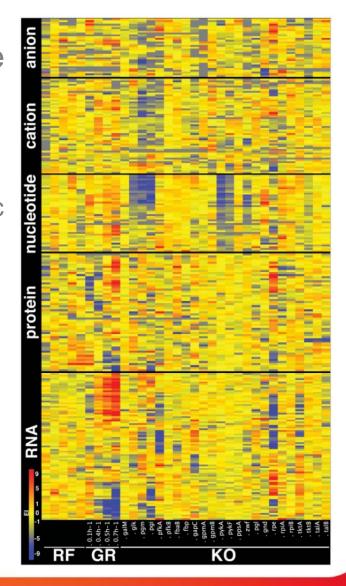




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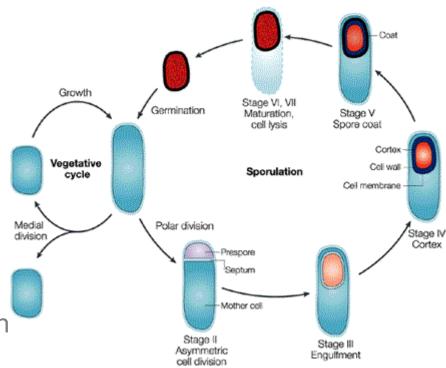
Sporulation and cell differentiation

 Bacillus subtilis cells can form resistant spores when environmental conditions become unfavorable (starvation)

Ultimate response in repertoire of stress responses (motility, toxin release, competence, ...)

 Asymmetric cell division produces smaller forespore cell and larger mother cell

Prototype of cell differentiation and intercellular signalling



Errington (2005), Nat. Rev. Microbiol., 1(2):117-126

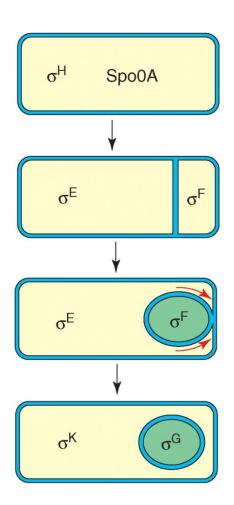




Sporulation and cell differentiation

- Precise temporal ordering of events on molecular level
- Specific proteins control differentiation processes in different stages of sporulation

Transcription factors, sigma factors



Piggot and Hilbert (2004), Curr. Opin. Microbiol., 7(6):579-86





General questions on cellular adaptation

 Cells are capable of responding to a variety of changes in their environment by adapting their physiology

Change in carbon source, starvation, population density, ...

 On the molecular level, these responses involve adjustment of protein concentrations in the cell

Enzymes, sigma factors, transcription factors, ...

- Question: how can protein concentrations change in response to specific environmental changes?
- Question: how does cell coordinate changes in concentration of a variety of proteins?
- Changes in protein concentrations involve changes in gene expression

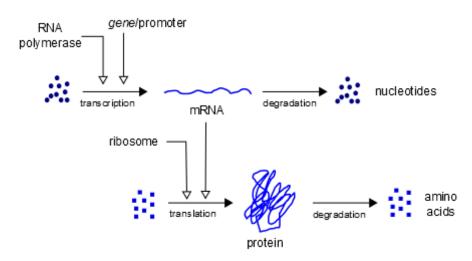




Gene expression

- Typically, and simplifying quite a bit, gene expression in bacteria involves:
 - Transcription by RNA polymerase (mRNA)
 - Translation by ribosomes (proteins)
 - Degradation of mRNA and protein

Biochemical view:



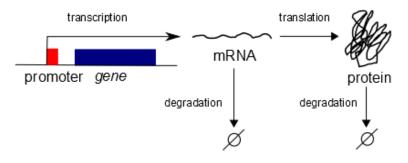




Gene expression

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 - Transcription by RNAP polymerase (mRNA)
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Simplified view:

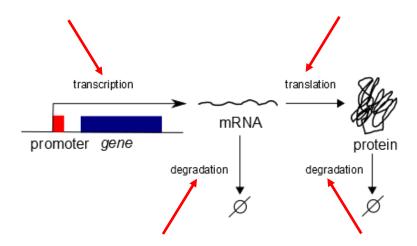






Regulation of gene expression

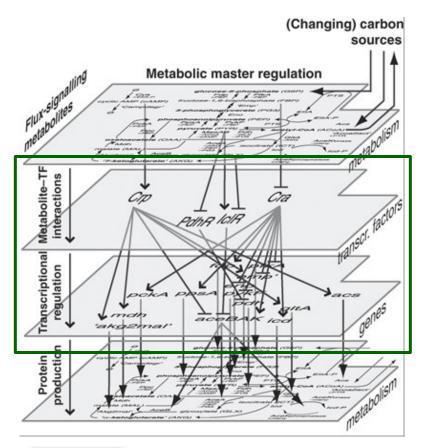
- Typically, and simplifying quite a bit, regulation of gene expression in bacteria involves:
 - Transcription regulation by transcription factors
 - Translation regulation by small RNAs
 - Regulation of degradation by proteases







 Gene regulatory networks control changes in gene expression levels in response to environmental perturbations



 Gene regulatory networks consist of genes, gene products, signalling metabolites, and their mutual regulatory interactions

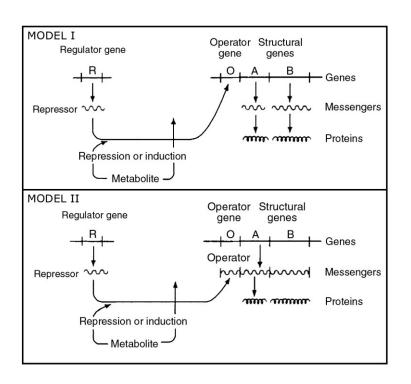
Global regulators of transcription involved in glucose-acetate diauxie in *E. coli*

Kotte et al. (2010), Mol. Syst. Biol., 6:355





 Gene regulatory networks control changes in expression levels in response to environmental perturbations



 Gene regulatory networks consist of genes, gene products, signalling metabolites, and their mutual regulatory interactions

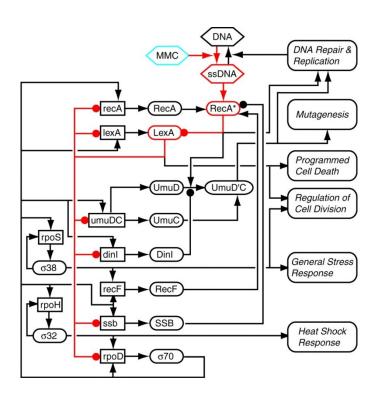
Original lac operon model

Jacob and Monod (1961), J. Mol. Biol., 3(3):318-56





 Gene regulatory networks control changes in expression levels in response to environmental perturbations



 Gene regulatory networks consist of genes, gene products, signalling metabolites, and their mutual regulatory interactions

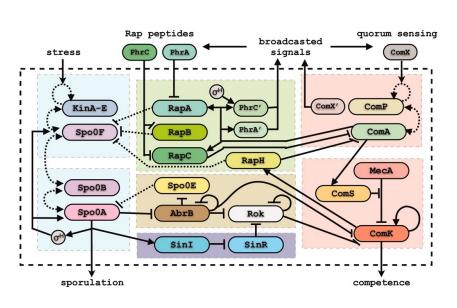
SOS response network in E. coli

Gardner et al. (2011), Science, 301(5629):102-5





 Gene regulatory networks control changes in expression levels in response to environmental perturbations



Gene regulatory networks consist of genes, gene products, signalling metabolites, and their mutual regulatory interactions

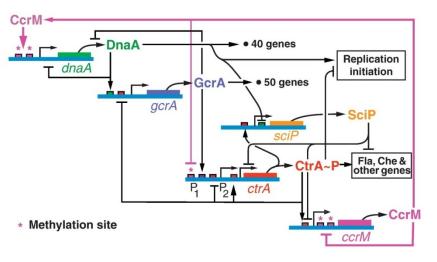
Sporulation and competence network in *B. subtilis*

Schultz et al. (1961), Proc. Natl. Acad. Sci. USA, 106(50):21027-34





 Gene regulatory networks control changes in expression levels in response to environmental perturbations



Gene regulatory networks consist of genes, gene products, signalling metabolites, and their mutual regulatory interactions

Cauleobacter cell cycle network

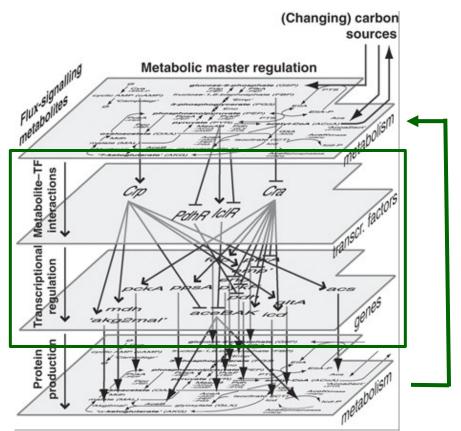
McAdams and Shapiro (2011), J. Mol. Biol., 409(1):28-35





Broader view on gene regulatory networks

 Gene regulatory networks control changes in expression levels in response to environmental perturbations



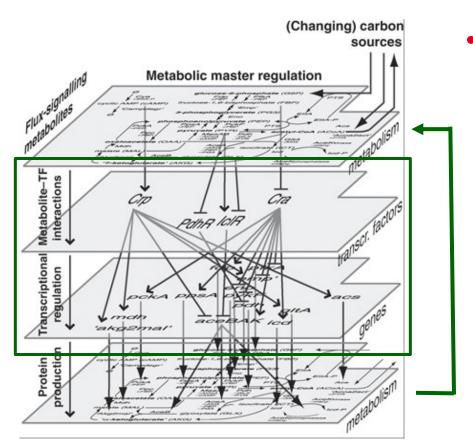
- But: adaptation of gene expression leads to changes in metabolism which feed back into regulatory network
- Gene regulatory networks are intertwined with metabolic and signaling networks
 - Complex, heterogeneous systems evolving on different time-scales





Broader view on gene regulatory networks

 Gene regulatory networks control changes in expression levels in response to environmental perturbations



 Feedback through metabolism leads to indirect regulatory interactions: metabolic coupling

Regulatory effects of enzymes on gene expression

Baldazzi et al. (2010), PLoS Comput. Biol., 6(6):e1000812

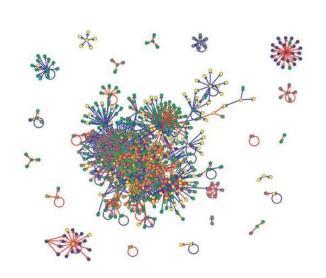




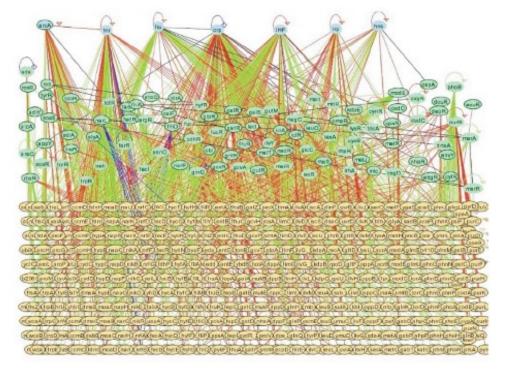
Complexity of gene regulatory networks

 Most gene regulatory networks of biological interest are large and complex

E. coli has 4200 genes coding for several hundreds of transcription factors



Cases and de Lorenzo (2005), *Nat. Rev. Microbiol.*, 3(2):105-18



Martinez-Antonio et al. (2003), Curr. Opin. Microbiol., 6(5):482-9





Systems biology

- Most gene regulatory networks of biological interest are large and complex
- No global view of functioning of network available, despite abundant knowledge on network components
 - Understanding of dynamics requires **experimental tools** for monitoring gene expression over time
 - Understanding of dynamics requires **mathematical modeling** and **computer analysis and simulation**
 - Discipline now often referred to as systems biology

Alon (2007), An Introduction to Systems Biology, Chapman & Hall/CRC Press





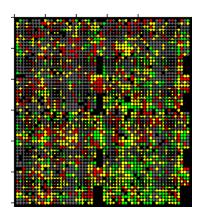
Experimental tools

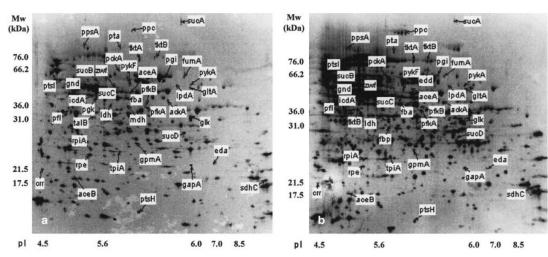
 A variety of experimental tools allow gene expression to be measured, by quantifying mRNA and protein abundancies

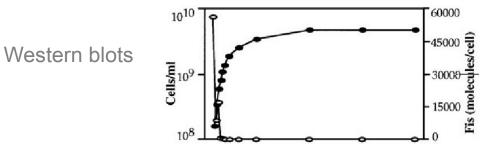
Peng and Shimizu (2003), *App. Microbiol. Biotechnol.*, 61:163-78

2D gels

DNA microarrays







Ali Azam et al. (1999), J. Bacteriol., 181(20):6361-70

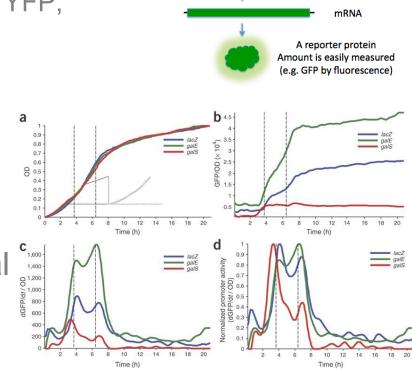




Fluorescent reporter genes

- Use of fluorescent reporter genes allows expression from host promoter to be monitored in vivo and in real time
 - Different colors (emission peaks): GFP, YFP, RFP, ...
 - Reporter genes on plasmids and on chromosome
 - Transcriptional and translational reporters
- Library of fluorescent transcriptional reporter genes in *E. coli*

Zaslaver et al. (2006), Nat. Methods, 3(8):623-8



Reporter gene (e.g. encoding GFP or

luciferase)

Regulatory sequence to

be studied

(e.g. a gene's promoter)



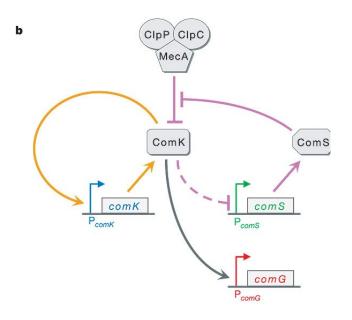


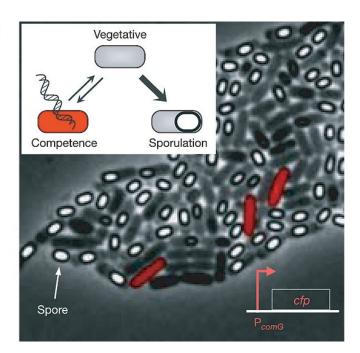
DNA

Single-cell microscopy

- Monitoring of gene expression in single cells using fluorescent reporters, automated time-lapse microscopy, and image analysis
- Monitoring onset of competence in B. subtilis

Süel et al. (2006), Nature, 440:545-50



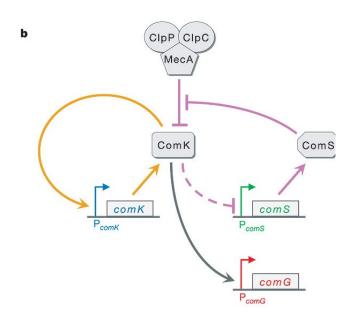


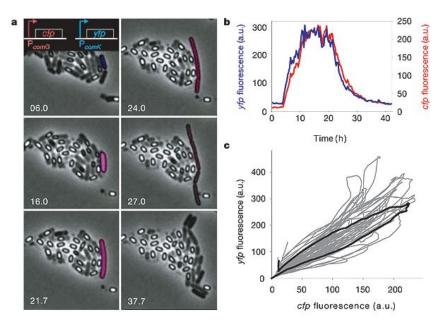




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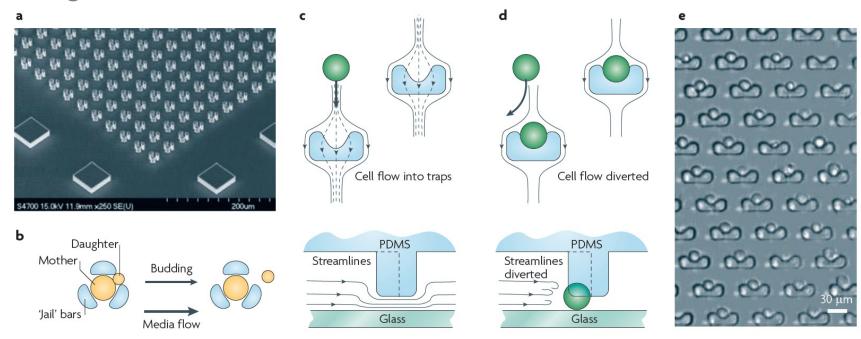






Single-cell microscopy and microfluidics

 Microfluidic trapping devices for long-term acquisition of single-cell data



Microfluidic devices allow tight control of environmental perturbations

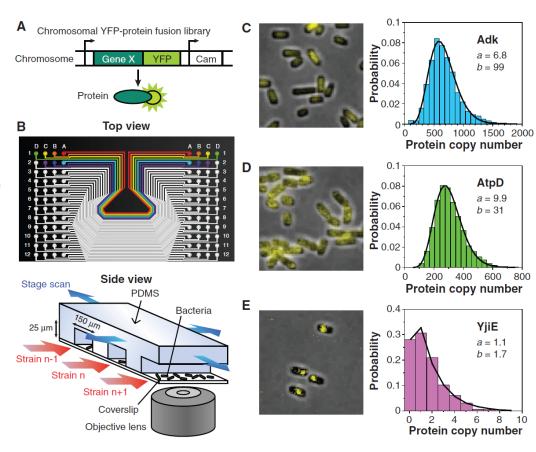
Bennett and Hasty (2009), Nat. Rev. Genet., 10(9):628-38





Single-molecule quantification

- expression at singlemolecule level using
 fluorescence reporter
 genes, microfluidic device,
 fluorescence microscopy,
 and calibration
- Measurement of expression of thousand *E.* coli genes using YFPtagged chromosomal reporters



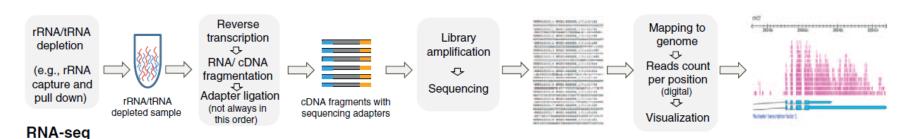
Taniguchi et al. (2010), Science, 329(5991):533-9





RNA sequencing

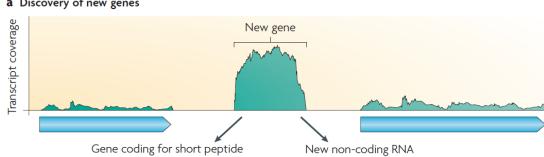
 RNA sequencing (RNA-seq) exploits new generation of sequencing technologies for quantifying RNA levels



Mäder et al. (2010), Curr. Opin. Biotechnol., 22(1):32-41

Use of RNA-seq data to discover new genes and detect operon structure
 a Discovery of new genes

Sorek and Cossart (2010), *Nat. Rev. Genet.*, 11(1):9-16

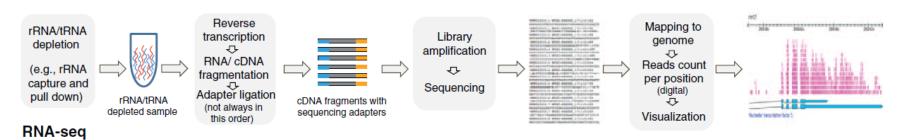






RNA sequencing

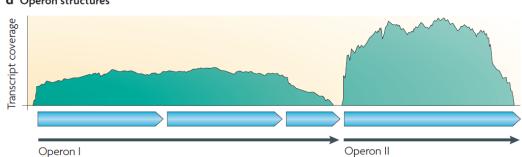
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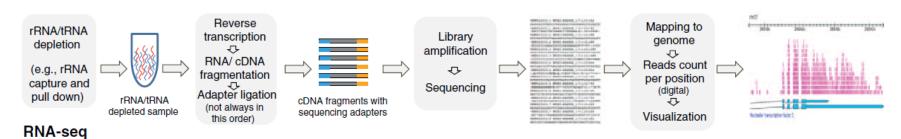






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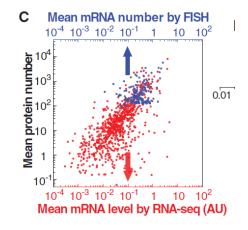
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 Use of RNA-seq to determine correlation between mean RNA and mean protein levels

Taniguchi et al. (2010), Science, 329(5991):533-9



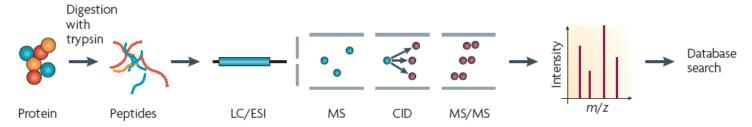




Quantitative proteomics

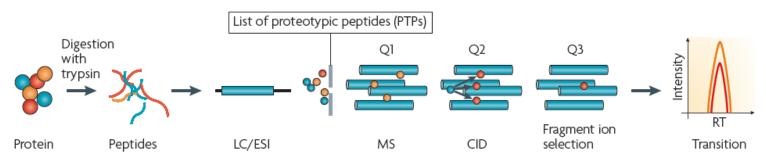
 Measurement of protein abundance using mass-spectrometrybased techniques (quantitative proteomics)

Use of calibration standards to achieve absolute quantification



Gstaiger and Aebersold (2009), Nat. Rev. Genet., 10:617-27

 Several targeted proteomics techniques developed to improve quantification of low-abundance proteins

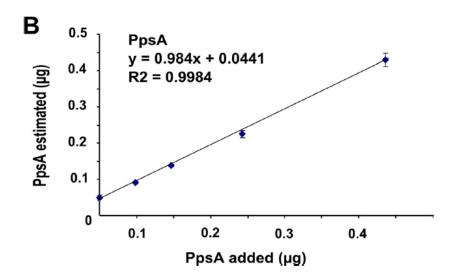


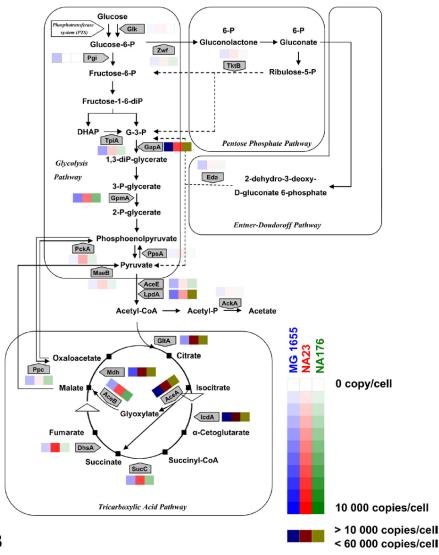




Quantitative proteomics

 Absolute quantification of proteins in central metabolism in *E. coli*





Trachessec et al. (2014), Mol. Cell. Proteom., 13(4):954-68





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- Most gene regulatory networks of biological interest are large and complex
- No global view of functioning of network available, despite abundant knowledge on network components
 - Understanding of dynamics requires **experimental tools** for monitoring gene expression over time
 - Understanding of dynamics requires **mathematical modeling** and **computer analysis and simulation**
 - Discipline now often referred to as systems biology

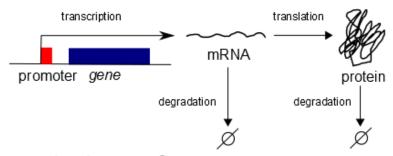
Alon (2007), An Introduction to Systems Biology, Chapman & Hall/CRC Press





Modeling of gene regulatory networks

 Modeling of gene regulatory networks amount to modeling of gene expression and regulation of gene expression



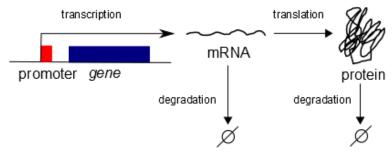
- Possible aims of modeling of gene regulatory networks:
 - Understanding role of individual components and interactions
 - Suggesting missing components and interactions
- Advantages of mathematical and computer tools:
 - Precise and unambiguous description of network
 - Systematic derivation of predictions of network behavior





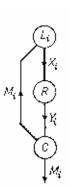
Modeling of gene regulatory networks

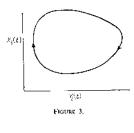
 Modeling of gene regulatory networks amount to modeling of gene expression and regulation of gene expression

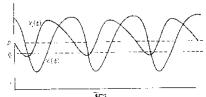


 First models of gene regulatory networks date back to early days of molecular biology

Feedback circuits and oscillators







Goodwin (1963), Temporal Organization in Cells





Modeling of gene regulatory networks

 Different modeling formalisms exist, describing gene expression on different levels of detail



Stochastic master equations

Ordinary differential equations (ODEs)

Boolean networks

Smolen et al. (2000), Bull. Math. Biol., 62(2):247-292
Hasty et al. (2001), Nat. Rev. Genet., 2(4):268-279
de Jong (2002), J. Comput. Biol., 9(1): 69-105
Szallassi et al. (2006), System Modeling in Cellular Biology, MIT Press
Bolouri (2008), Computational Modeling of Gene Regulatory Networks, Imperial College Press
Karleback and Shamir (2008), Nat. Rev. Mol. Cell Biol., 9(10):770-80





Conclusions

- Gene regulatory networks control adaptive response of bacteria to changes in environment
- Gene regulatory networks are intertwined with metabolic and signaling networks
- Technology for measuring gene expression over time, and thus functioning of gene regulatory networks, are rapidly developing
- Modeling necessary for understanding dynamics of complex networks: systems biology
- A variety of formalisms for modeling gene regulatory networks, in a detailed or coarse-grained way, have been developed





Merci!



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