

Qualitative modeling of gene regulatory networks

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INRIA Grenoble - Rhône-Alpes and IBIS



- IBIS: systems biology group at INRIA/Université Joseph Fourier/CNRS
 - Analysis of bacterial regulatory networks by means of models and experiments
 - Biologists, computer scientists, mathematicians, physicists, ...

http://team.inria.fr/ibis







Overview

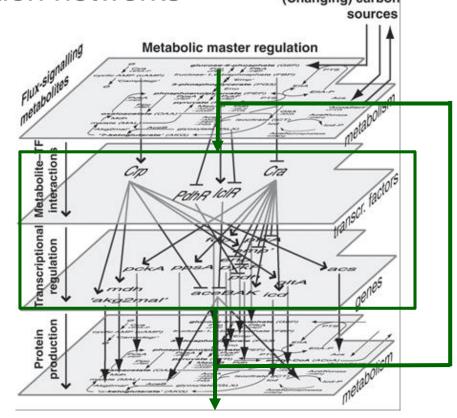
- Part 1. Systems biology and kinetic modeling
- Part 2. Metabolic network modeling
- Part 3. Gene regulatory network modeling
 - Quantitative modeling of gene regulatory networks
 - Qualitative modeling of gene regulatory networks
 - Stochastic modeling of gene regulatory networks
 - Practical on integrated models of bacterial growth (Matlab)
- Part 4. Models and data





Gene regulatory networks

- Focus on **subsystems** that can be studied in isolation due to **modular structure** of reaction networks (Changing) carbon
 - Time-scale hierarchies
 - Connectivity structure
- Gene regulatory networks
 - Genes, proteins, and regulatory interactions
 - Reactions involved in transcription and translation and their regulation
 - Time-scale: min (mRNA) to h (proteins)



Kotte et al. (2010), Mol. Syst. Biol., 6: 355





Modeling of gene regulatory networks

 Well-established theory for modeling of gene regulatory networks using ordinary differential equation (ODE) models

Bolouri (2008), Computational Modeling of Gene Regulatory Networks, Imperial College Press Polynikis et al. (2009), J. Theor. Biol., 261(4):511-30

- Practical problems encountered by modelers:
 - Knowledge on molecular mechanisms rare
 - Quantitative information on kinetic parameters and molecular concentrations absent
 - Large models





Qualitative modeling and simulation

- Intuition: essential properties of network dynamics robust against reasonable model simplifications
- Qualitative modeling and simulation of large and complex gene regulatory networks using simplified ODE models

de Jong, Gouzé et al. (2004), Bull. Math. Biol., 66(2):301-40

 Close proximity with discrete, logical models of gene regulation

Thomas and d'Ari (1990), *Biological Feedback*, CRC Press Kauffman (1993), *The Origins of Order*, Oxford University Press



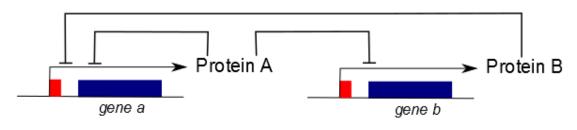


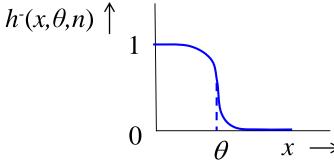
Ordinary differential equation models

 Gene regulatory networks modeled by ODE models using sigmoid functions to describe regulatory interactions

$$\dot{x}_a = \kappa_a h^{\text{-}}(x_a, \theta_{a2}, n) h^{\text{-}}(x_b, \theta_b, n) - \gamma_a x_a$$

$$\dot{x}_b = \kappa_b h^{\text{-}}(x_a, \theta_{a1}, n) - \gamma_b x_b$$





x : protein concentration

 θ : threshold concentration

 κ , γ : rate constants

n : steepness parameter

 Expressions of sigmoid functions account for combinatorial control of gene expression (AND, OR, NOR, ...)



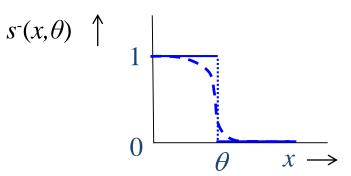


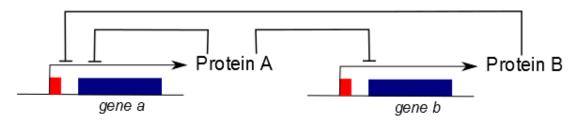
PL differential equation models

 ODE models approximated by means of step functions to describe regulatory interactions

$$\dot{x}_a = \kappa_a \, s^{\scriptscriptstyle -}(x_a, \theta_{a2}) \, s^{\scriptscriptstyle -}(x_b, \theta_b) - \gamma_a \, x_a$$

$$\dot{x}_b = \kappa_b \, s^{\scriptscriptstyle -}(x_a, \theta_{a1}) - \gamma_b \, x_b$$





x : protein concentration

 θ : threshold concentration

 κ , γ : rate constants

Piecewise-linear (PL)DE models of gene regulatory networks

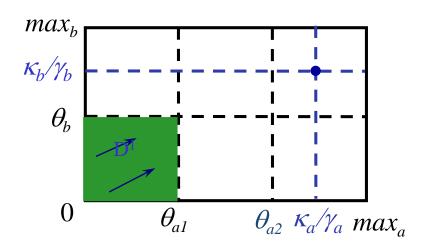
Glass and Kauffman (1973), J. Theor. Biol., 39(1):103-29





Analysis of local dynamics of PL models

Monotone convergence towards focal point in regions separated by thresholds



$$\dot{x}_a = \kappa_a - \gamma_a \, x_a$$

$$\dot{x}_b = \kappa_b - \gamma_b \, x_b$$

$$\dot{x}_a = \kappa_a \, s^{\text{-}}(x_a, \theta_{a2}) \, s^{\text{-}}(x_b, \theta_b) - \gamma_a \, x_a$$
$$\dot{x}_b = \kappa_b \, s^{\text{-}}(x_a, \theta_{a1}) - \gamma_b \, x_b$$

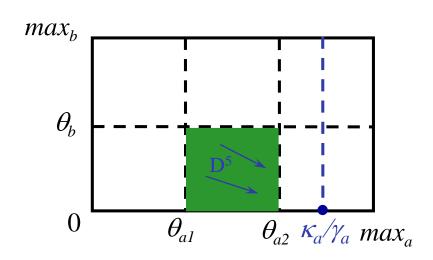
Glass and Kauffman (1973), J. Theor. Biol., 39(1):103-29





Analysis of local dynamics of PL models

Monotone convergence towards **focal point** in regions separated by thresholds



$$\dot{x}_a = \kappa_a - \gamma_a x_a$$

$$\dot{x}_b = -\gamma_b x_b$$

$$\dot{x}_a = \kappa_a \, s^{\text{-}}(x_a, \theta_{a2}) \, s^{\text{-}}(x_b, \theta_b) - \gamma_a \, x_a$$
$$\dot{x}_b = \kappa_b \, s^{\text{-}}(x_a, \theta_{a1}) - \gamma_b \, x_b$$

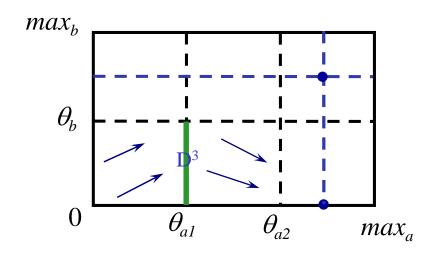
Glass and Kauffman (1973), J. Theor. Biol., 39(1):103-29





Analysis of local dynamics of PL models

Instantaneous crossing of regions located on thresholds, or ...



$$\dot{x}_a = \kappa_a \, s^{\scriptscriptstyle -}(x_a, \theta_{a2}) \, s^{\scriptscriptstyle -}(x_b, \theta_b) - \gamma_a \, x_a$$

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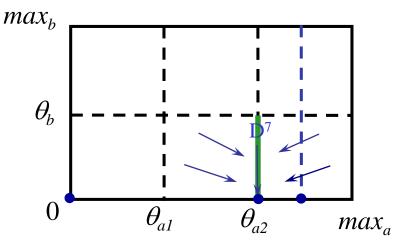




Analysis of local dynamics of PL models

... quasi-monotone convergence towards focal sets located on threshold

hyperplanes



$$\dot{x}_a = \kappa_a \, s^{\text{-}}(x_a, \theta_{a2}) \, s^{\text{-}}(x_b, \theta_b) - \gamma_a \, x_a$$
$$\dot{x}_b = \kappa_b \, s^{\text{-}}(x_a, \theta_{a1}) - \gamma_b \, x_b$$

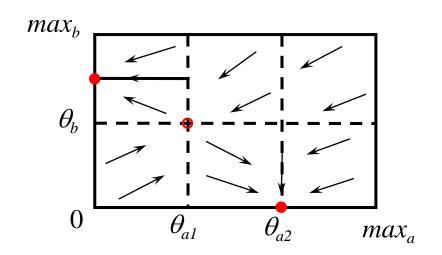
Extension of PL differential equations to differential inclusions
using Filippov approach
 Gouzé and Sari (2002), Dyn. Syst., 17(4):299-316

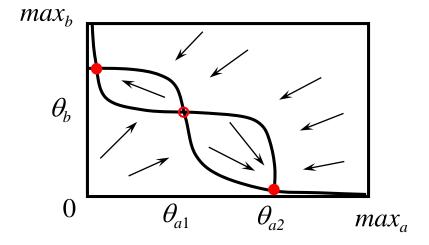




 Analysis of global dynamics obtained by piecing together local dynamics in regions

PL approximation preserves bistability of cross-inhibition network



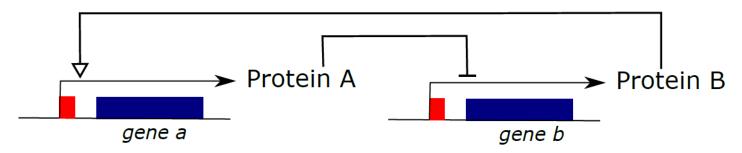






Simple oscillator network

 Question: write out the PLDE model for a simple oscillator network



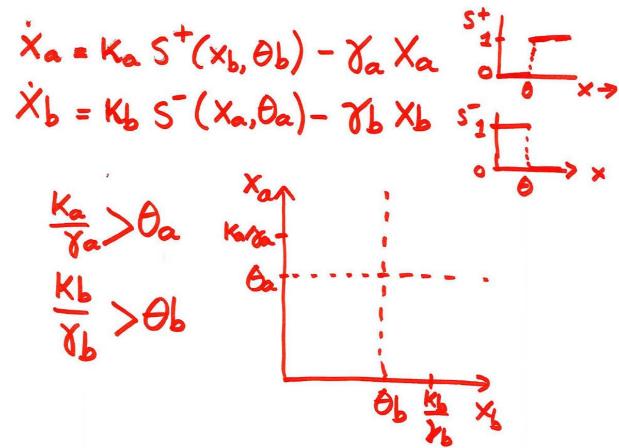
Question: perform phase space analysis of PLDE model





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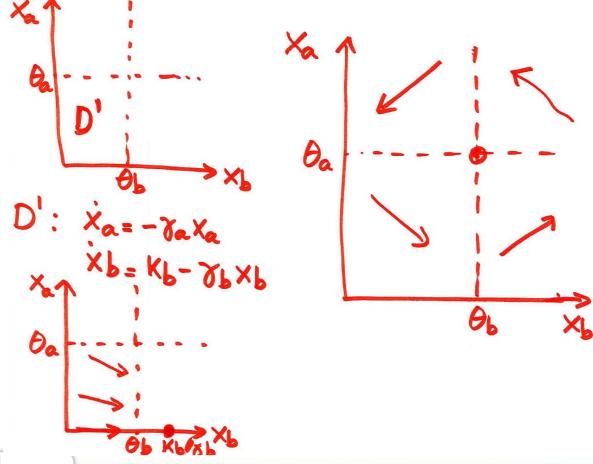






Simple oscillator network

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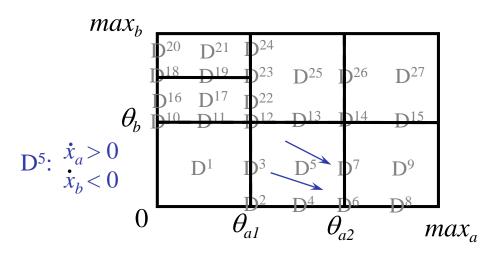




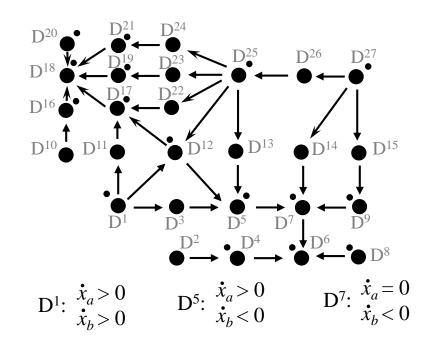


- State space can be partitioned into regions with unique derivative sign pattern
- Qualitative abstraction yields state transition graph that provides discrete picture of continuous dynamics

Alur et al. (2000), Proc. IEEE, 88(7):971-84



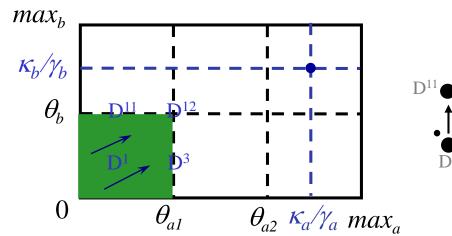
de Jong et al. (2004), Bull. Math. Biol., 66(2):301-40 Batt et al. (2008), Automatica, 44(4):982-9

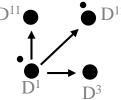






- State transition graph gives conservative approximation of continuous dynamics
 - Every solution of PL model corresponds to path in state transition graph
 - Converse is not necessarily true!
- State transition graph is invariant for given inequality constraints on parameters





$$0 < \theta_{a1} < \theta_{a2} < \kappa_a / \gamma_a < max_a$$
$$0 < \theta_b < \kappa_b / \gamma_b < max_b$$

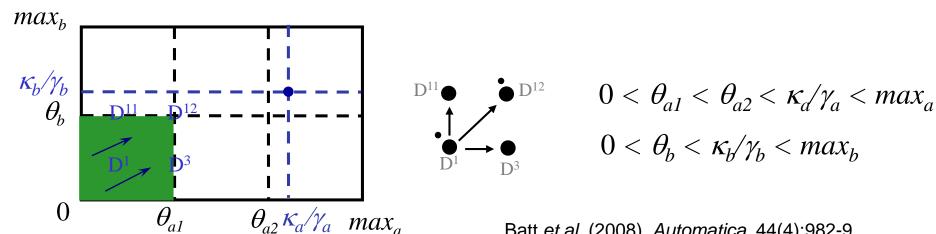
$$0 < \theta_b < \kappa_b/\gamma_b < max_b$$

Batt et al. (2008), Automatica, 44(4):982-9





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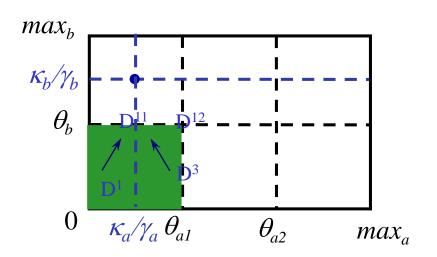


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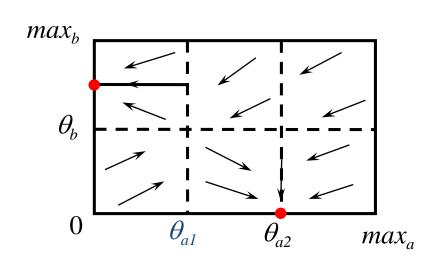


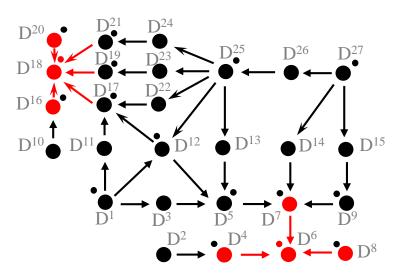


Use of state transition graph

- Analysis of steady states and limit cycles of PL models
 - Attractor states in graph correspond (under certain conditions) to
 stable steady states of PL model Casey et al. (2006), J. Math Biol., 52(1):27-56
 - Attractor cycles in graph correspond (under certain conditions) to stable limit cycles of PL model

Glass and Pasternack (1978), *J. Math Biol.*, 6(2):207-23 Edwards (2000), *Physica D*, 146(1-4):165-99



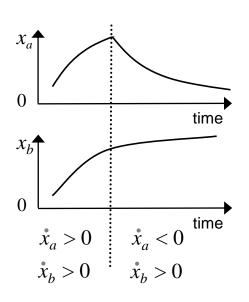




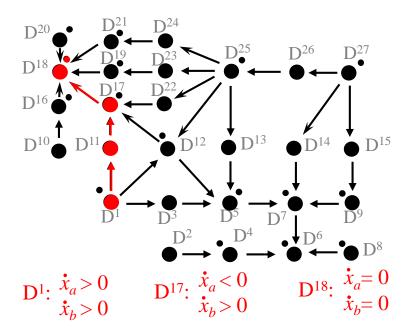


Use of state transition graph

- Paths in state transition graph represent predicted sequences of qualitative events
- Model validation: comparison of predicted and observed sequences of qualitative events



Concistency?
Yes



Need for automated and efficient tools for model validation





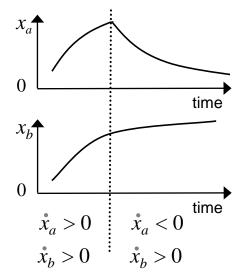
Model validation by model checking

Dynamic properties of system can be expressed in temporal

logic (CTL)

There Exists a Future state where $\dot{x}_a > 0$ and $\dot{x}_b > 0$ and starting from that state, there Exists a Future state where $\dot{x}_a < 0$ and $\dot{x}_b > 0$

$$EF(\dot{x}_a > 0 \land \dot{x}_b > 0 \land EF(\dot{x}_a < 0 \land \dot{x}_b > 0))$$



 Model checking is automated technique for verifying that state transition graph satisfies temporal-logic statements

Efficient computer tools available for model checking

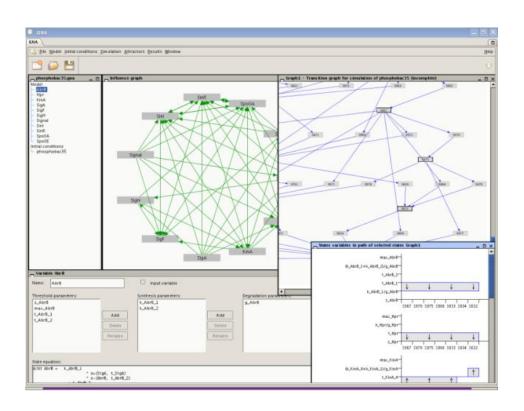
Batt et al. (2005), Bioinformatics, 21(supp. 1): i19-i28





Genetic Network Analyzer (GNA)

 Qualitative analysis of PL models implemented in Java: Genetic Network Analyzer (GNA)





de Jong *et al.* (2003), *Bioinformatics*, 19(3):336-44 http://www-helix.inrialpes.fr/gna

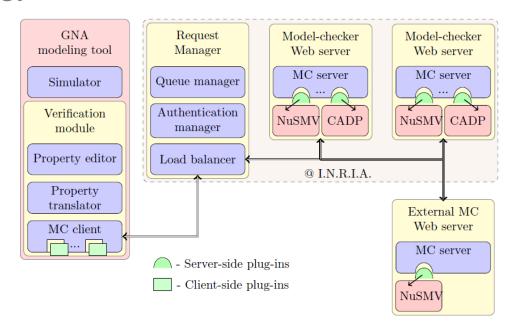


Genetic Network Analyzer (GNA)

- Model-checking technology made available to GNA user
 - Develop temporal logics tailored to biological questions

Mateescu et al. (2011), Theor. Comput. Sci., 412:2854-83

Develop temporal-logic patterns for frequently-asked modeling questions



Monteiro et al. (2008), Bioinformatics, 24(16):i227-33

Connect GNA to standard model checkers through a web-server connection

Monteiro et al., (2009), BMC Bioinform., 10:450





Analysis of bacterial regulatory networks

- Applications of qualitative simulation in bacteria:
 - Initiation of sporulation in Bacillus subtilis

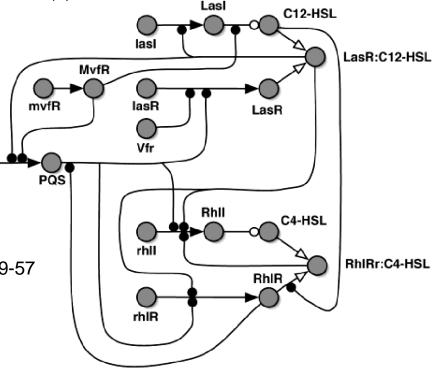
de Jong, Geiselmann et al. (2004), Bull. Math. Biol., 66(2):261-300

 Quorum sensing in Pseudomonas aeruginosa

Viretta and Fussenegger (2004), *Biotechnol. Prog.*, 20(3):670-8

 Onset of virulence in Erwinia chrysanthemi

Sepulchre et al. (2007), J. Theor. Biol., 244(2):239-57



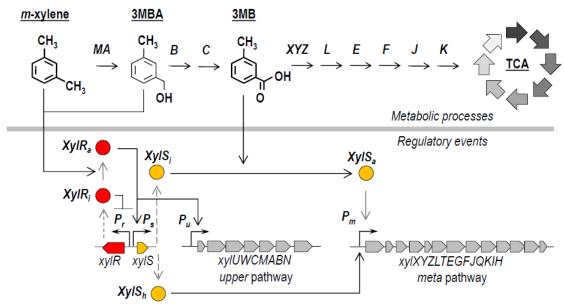




Biodegradation of polluants by P. putida

 Soil bacterium Pseudomonas putida mt-2 is archetypal model for environmental biodegradation of aromatic pollutants

TOL network involved in degradation of *m*-xylene to intermediates for central carbon metabolism



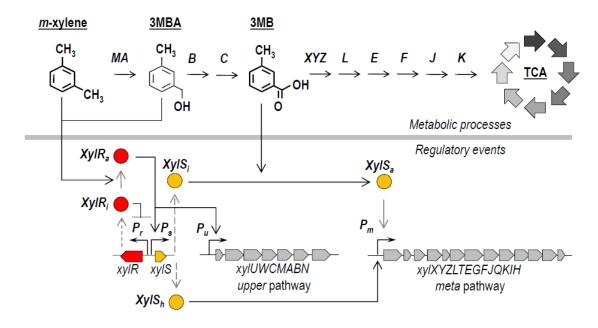
Rocha-Silva et al. (2011), Environ. Microbiol., 13(9):2389-402





Role of regulators of TOL network

 Question: what is the role of the central, plasmid-encoded regulators XyIR and XyIS?



Development of PL model of TOL network

Translation of network diagram into regulatory logic and PL model

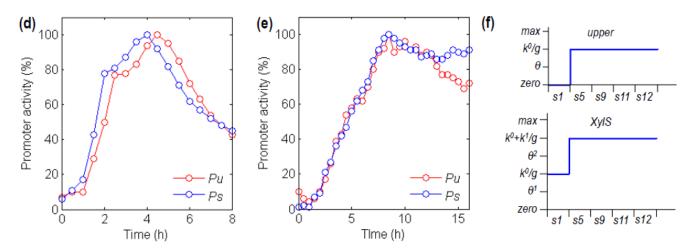
Rocha-Silva et al. (2011), BMC Syst. Biol., 5:191





Role of regulators of TOL network

• Validation of model by testing predictions under different perturbation conditions (mutants, metabolic inducers, ...)



Plasmid-encoded regulators of TOL network act as regulatory firewall

Prevent toxic *m*-xylene and its biodegradation intermediates from intervening with indigenous metabolic pathways

Rocha-Silva et al. (2011), BMC Syst. Biol., 5:191



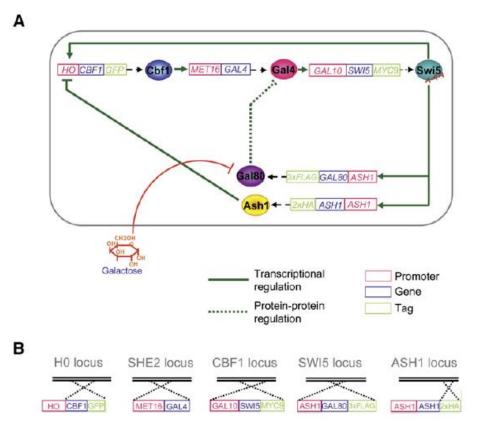


IRMA: synthetic network in yeast

IRMA: synthetic network in yeast consisting of interlocked positive and negative feedback loops

Networks functions independently from host cell

 Network can be externally controlled by growing cells in glucose or galactose



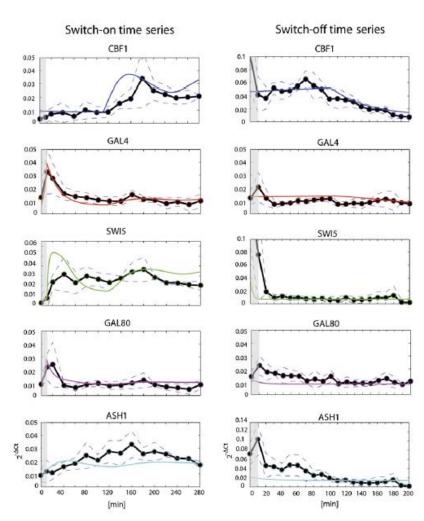
Cantone et al. (2009), Cell, 137(1):172-81





IRMA: synthetic network in yeast

- IRMA proposed as a benchmark for modeling and identification approaches
- IRMA dynamics measured over time in galactose (switch-on) and glucose (switch-off)
 Quantitative RT-PCR
- Question: are measured dynamics consistent with constructed network structure?

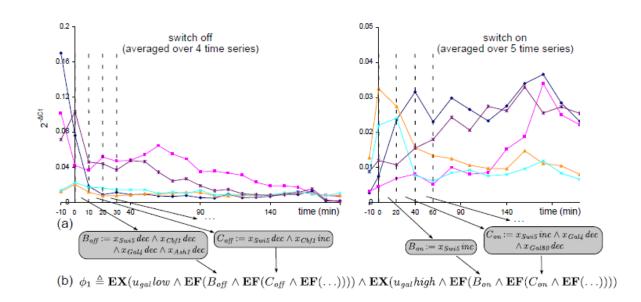


Cantone et al. (2009), Cell, 137(1):172-81





- Development of (unparametrized) PL model representing network structure
- Approach to test consistency between network structure and data based on automated parameter constraint search:
 - Generate temporal logic formulae encoding observed network dynamics



Batt et al. (2010), Bioinformatics, 26(18):i603-10





- Development of (unparametrized) PL model representing network structure
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 - Generate temporal logic formulae encoding observed network dynamics
 - Test if there are any parametrizations of PL model satisfy temporal logic formulae

		Symbolic state space and symbolic parameter space		Symbolic state space and explicit parameter space	
Γ	Property	Existence of	Parametrization*	Number of	Parametrization*
L		parametrization		parametrizations	
	ϕ_1 : averaged time-series	Yes (49 s)	$\frac{\kappa_{Swi5}^{0}}{\gamma_{Swi5}} < \theta_{Swi5}^{g} < \theta_{Swi5}^{c} < \theta_{Swi5}^{a} < \frac{\kappa_{Swi5}^{0} + \kappa_{Swi5}}{\gamma_{Swi5}}$ $\wedge \frac{\kappa_{Gal80}^{0}}{\gamma_{Gal80}} < \frac{\kappa_{Gal80}^{0} + \kappa_{Gal80}}{\gamma_{Gal80}} < \theta_{Gal80}$	(925 s)	$\frac{\kappa_{Swi5}^{0}}{\gamma_{Swi5}} < \theta_{Swi5}^{c} < \theta_{Swi5}^{a} < \frac{\kappa_{Swi5}^{0} + \kappa_{Swi5}}{\gamma_{Swi5}} \land$ $(\theta_{Gal80} < \frac{\kappa_{Gal80}^{0}}{\gamma_{Gal80}} \land \frac{\kappa_{Swi5}^{0}}{\gamma_{Swi5}} < \theta_{Swi5}^{g} < \frac{\kappa_{Swi5}^{0} + \kappa_{Swi5}}{\gamma_{Swi5}}$ $\lor \frac{\kappa_{Gal80}^{0}}{\gamma_{Gal80}} < \theta_{Gal80} < \frac{\kappa_{Gal80}^{0} + \kappa_{Gal80}}{\gamma_{Gal80}} \land \frac{\kappa_{Swi5}^{0}}{\gamma_{Swi5}} < \theta_{Swi5}^{g}$ $\lor \frac{\kappa_{Gal80}^{0} + \kappa_{Gal80}}{\gamma_{Gal80}} < \theta_{Gal80})$

^{*}All parametrizations additionally include $\kappa^1_{Cbf1}/\gamma_{Cbf1} < \theta_{Cbf1} < (\kappa^1_{Cbf1} + \kappa^2_{Cbf1})/\gamma_{Cbf1} \wedge \kappa^0_{Gal4}/\gamma_{Gal4} < \theta_{Gal4} < (\kappa^0_{Gal4} + \kappa_{Gal4})/\gamma_{Gal4} \wedge \kappa^0_{Ash1}/\gamma_{Ash1} < \theta_{Ash1} < (\kappa^0_{Ash1} + \kappa_{Ash1})/\gamma_{Ash1}$





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 - Analyze parametrizations for biological plausibility

« Activation threshold of CBF1 by Swi5 higher than activation threshold of ASH1 »: confirmed by independent experimental data

Batt et al. (2010), Bioinformatics, 26(18):i603-10





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 - Generate temporal logic formulae encoding observed network dynamics
 - Test if there are any parametrizations of PL model satisfy temporal logic formulae
 - Analyze parametrizations for biological plausibility
- Automated approach for testing consistency based on modelchecking techniques
 - Symbolic encoding of model, dynamics and properties to make problem feasible



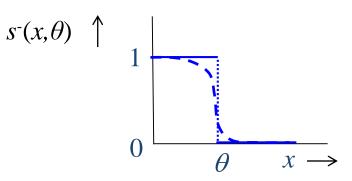


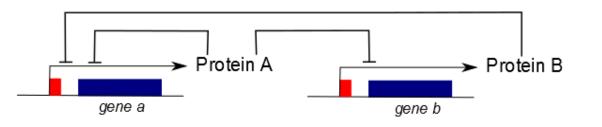
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x : protein concentration

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 Expressions of step functions account for combinatorial control of gene expression (AND, OR, NOR, ...)

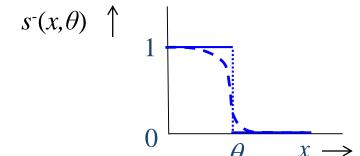


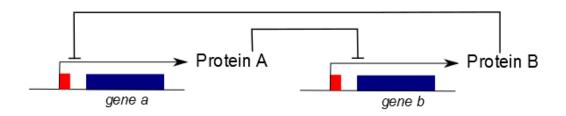


PL differential equation models

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Boolean models

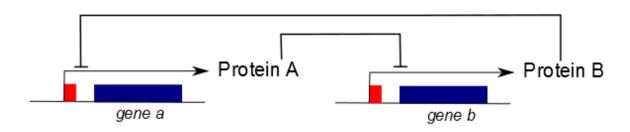
 Boolean models are discrete models of dynamics of gene regulatory networks

Thomas and d'Ari (1990), *Biological Feedback*, CRC Press Kauffman (1993), *The Origins of Order*, Oxford University Press Wang et al. (2012), *Phys. Biol.*, 9(5):055001

Boolean variables discretize state of gene regulatory network

$$X_a \in \{0, 1\}, X_b \in \{0, 1\}$$

 $X_a = (x_a > \theta_a), X_b = (x_b > \theta_b)$
 $X_a^t, X_b^t, t = 0, 1, 2, ...$







Boolean models

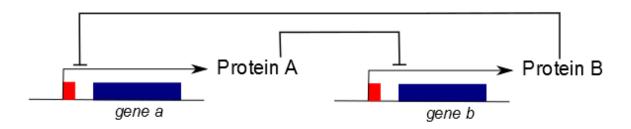
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- Boolean variables discretize state of gene regulatory network
- Boolean functions represent control of gene expression

$$X_a^{t+1} = \text{NOT } X_b^t$$

 $X_b^{t+1} = \text{NOT } X_a^t$

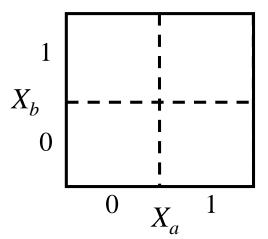






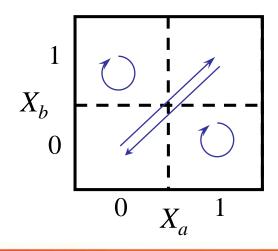
Analysis of Boolean models

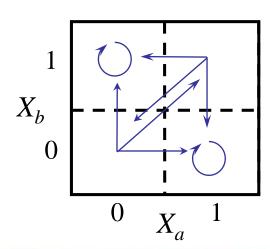
Boolean models can be analyzed in discrete state space



$$X_a^{t+1} = \text{NOT } X_b^t$$
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Synchrone and asynchrone dynamics



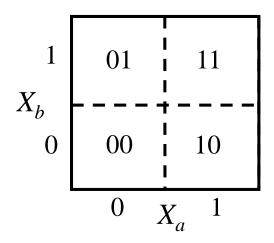


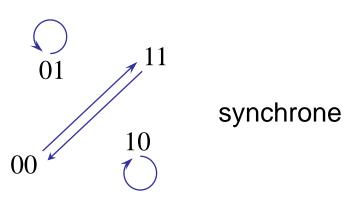


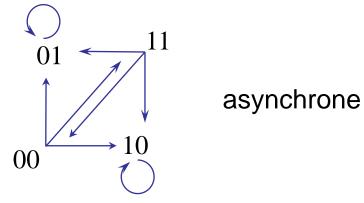


Analysis of Boolean models

- Dynamics of Boolean models can also be represented in state transition graph
 - Different graphs for synchrone and asynchrone dynamics
 - Attractors (states or cycles)











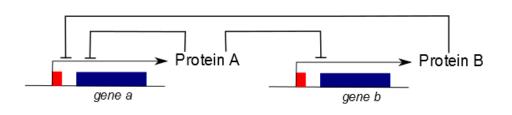
 Generalized logical models are discrete models of dynamics of gene regulatory networks

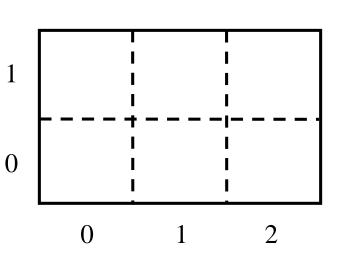
Thomas and d'Ari (1990), *Biological Feedback*, CRC Press Chaouiya *et al.* (2012), *Methods Mol. Biol.*, 804:463-79

Logical variables discretize state of gene regulatory network

$$X_a \in \{0, 1, 2, \ldots\}, X_b \in \{0, 1, 2, \ldots\}$$

$$X_a^t, X_b^t, t = 0, 1, 2, \dots$$







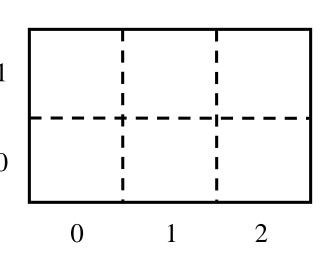


 Generalized logical models are discrete models of dynamics of gene regulatory networks

Thomas and d'Ari (1990), *Biological Feedback*, CRC Press Chaouiya *et al.* (2012), *Methods Mol. Biol.*, 804:463-79

- Logical variables discretize state of gene regulatory network
- Boolean functions represent control of gene expression

$$X_a^{t+1} = 2$$
, if $(X_a^t = 0 \text{ OR } X_a^t = 1) \text{ AND } X_b^t = 0$
 $X_a^{t+1} = 0$, if $X_a^t = 2 \text{ OR } X_b^t = 1$
 $X_b^{t+1} = 1$, if $X_a^t = 0$
 $X_b^{t+1} = 0$, if $X_a^t = 1 \text{ OR } X_a^t = 2$





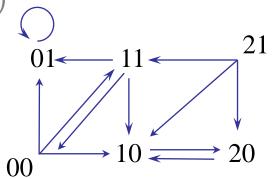


Generalized logical models are discrete models of dynamics of gene regulatory networks

Thomas and d'Ari (1990), Biological Feedback, CRC Press Chaouiya et al. (2012), Methods Mol. Biol., 804:463-79

- Logical variables discretize state of gene regulatory network
- Boolean functions represent control of gene expression
- Dynamics can be represented by state transition graph

Attractors (states and cycles)



asynchrone





 Generalized logical models are discrete models of dynamics of gene regulatory networks

Thomas and d'Ari (1990), *Biological Feedback*, CRC Press Chaouiya *et al.* (2012), *Methods Mol. Biol.*, 804:463-79

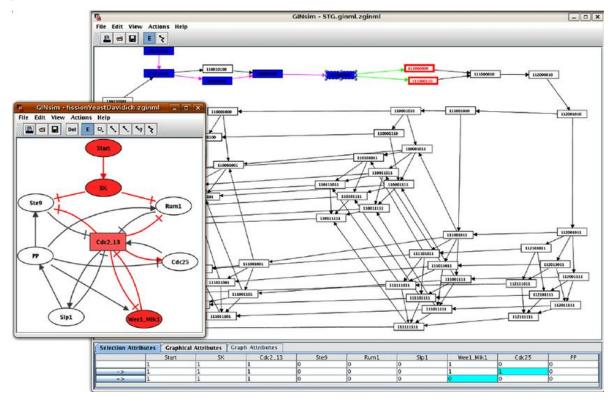
- Logical variables discretize state of gene regulatory network
- Boolean functions represent control of gene expression
- Dynamics can be represented by state transition graph
- Close correspondence between discrete abstractions of PLDE models and generalized logical models





GinSIM

GinSIM: computer tool for logical modeling of regulatory networks

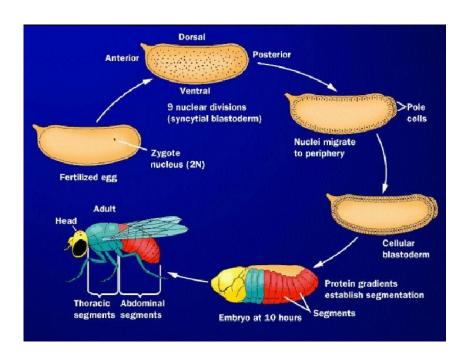


Chaouiya et al. (2012), Methods Mol. Biol., 804:463-79





Development of Drosophila melanogaster (fruit fly)



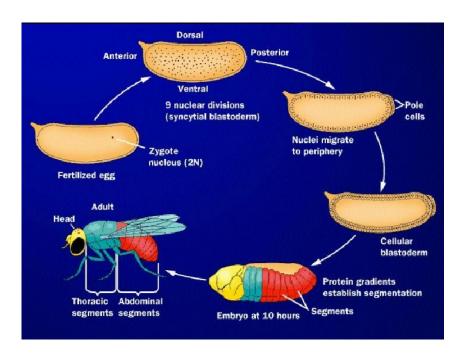


Purves et al. (1998), Life: The Science of Biology, Sinauer





Development of Drosophila melanogaster (fruit fly)

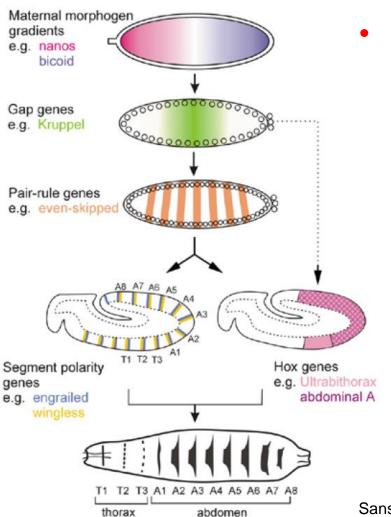


Purves et al. (1998), Life: The Science of Biology, Sinauer

Tomer et al. (2012), Nat. Methods, 9(7):755-63





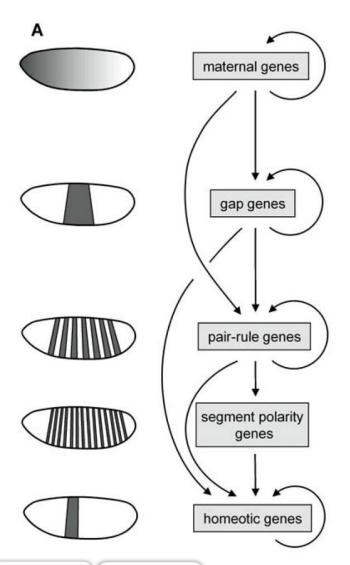


Spatiotemporal gene expression patterns during early development of *Drosophila* (fruit fly)

Sanson (2001), EMBO Rep., 2(12):1083-8





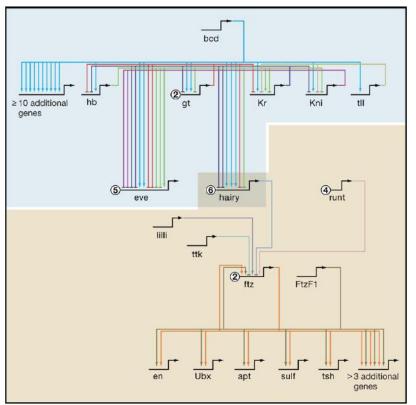


- Spatiotemporal gene expression patterns during early development of *Drosophila* (fruit fly)
- Gene classes and their interactions responsible for establishment of gene expression patterns

Schroeder et al. (2004), PLoS Biol., 4(2):e271







- Spatiotemporal gene expression patterns during early development of *Drosophila* (fruit fly)
- Gene classes and their interactions responsible for establishment of gene expression patterns
- Complex gene regulatory networks

Carroll (2008), Cell, 134(1):25-36

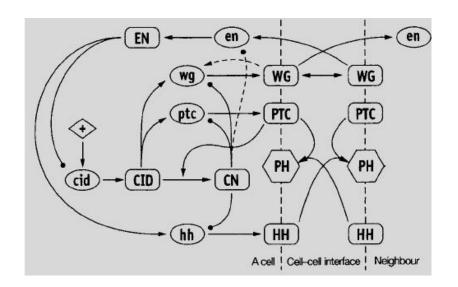


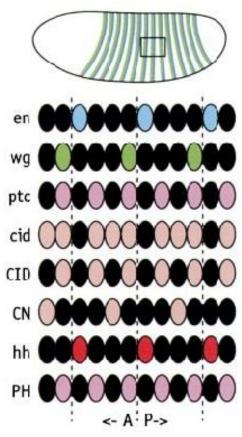


Model of *Drosophila* segmentation

Model of network of segment polarity genes in early development of Dresonhile

development of *Drosophila*





von Dassow et al. (2000), Nature, 406(6792): 188-92

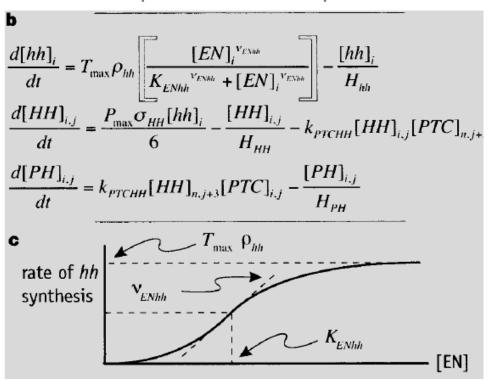


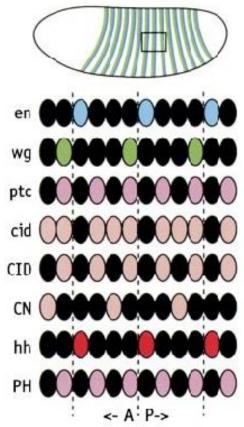


Model of *Drosophila* segmentation

 Model of network of segment polarity genes in early development of *Drosophila*

13 ODEs per cell and 48 parameters





von Dassow et al. (2000), Nature, 406(6792): 188-92



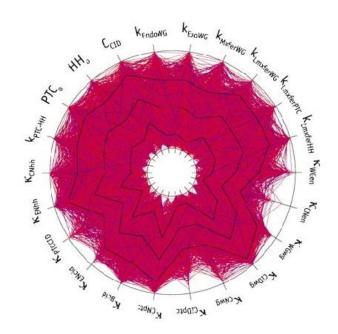


Robustness of gene expression patterns

 Spatial expression pattern of segment polarity genes robustly reproduced over large ranges of parameter values

0.5% of sampled parameter combinations leads to solution compatible

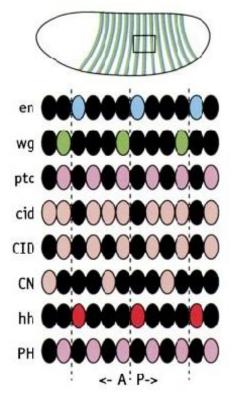
with data



von Dassow et al. (2000), Nature, 406(6792): 188-92

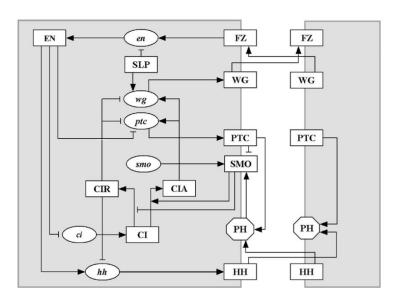






Logical model of *Drosophila* segmentation

 Logical model of segment polarity network: variables take values 0/1 and Boolean functions to update variables



Albert and Othmer (2003), J. Theor. Biol., 223(1):1-18

 hh_i HH_i ptc_i PTC_i PH_i

$$\begin{split} hh_i^{t+1} &= EN_i^t \text{ and not } CIR_i^t \\ HH_i^{t+1} &= hh_i^t \\ ptc_i^t &= CIA_i^{t+1} \text{ and not } EN_i^t \text{ and not } CIR_i^t \\ PTC_i^{t+1} &= ptc_i^t \text{ or } (PTC_i^t \text{ and not } HH_{i-1}^t \text{ and not } HH_{i+1}^t) \\ PH_i^t &= PTC_i^t \text{ and } (HH_{i-1}^t \text{ or } HH_{i+1}^t) \end{split}$$





Logical model of *Drosophila* segmentation

- Logical model of segment polarity network: variables take values 0/1 and Boolean functions to update variables
- Logical models are based on topology of network only (no parametrization), but are capable of reproducing experimental data: robustness

Observed initial state (before stage 8)

wg
WG
en
EN
hh
HH
ptc
PTC
PH
SMO
ci
CI
CIA
CIR

Albert and Othmer (2003), J. Theor. Biol., 223(1):1-18





Conclusions

- Modeling of genetic regulatory networks in bacteria often hampered by lack of information on parameter values
- Use of coarse-grained discrete or discretized models that provide reasonable approximation of dynamics
- Mathematical methods and computer tools for analysis of qualitative dynamics of discrete models
- Use of discrete models may gain insight into functioning of large and complex networks
- Discrete, coarse-grained models provide first idea of qualitative dynamics that may guide quantitative modeling





Merci!



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