

Hidde de Jong IBIS INRIA Grenoble – Rhône-Alpes Hidde.de-Jong@inria.fr

# INRIA Grenoble - Rhône-Alpes and IBIS



- IBIS: systems biology group at INRIA/Université Grenoble-Alpes
  - Analysis of bacterial regulatory networks by means of models and experiments
  - Biologists, computer scientists, mathematicians, physicists, ...

http://team.inria.fr/ibis







#### Overview

- Part 1. Systems biology and kinetic modeling
  - Introduction
  - Kinetic modeling of cellular reaction networks
- Part 2. Metabolic network modeling
  - Kinetic modeling of metabolism
  - Metabolic control analysis (MCA)
  - Flux balance analysis (FBA)
  - Practical on flux balance analysis (COBRA)
- Part 3. Gene regulatory network modeling



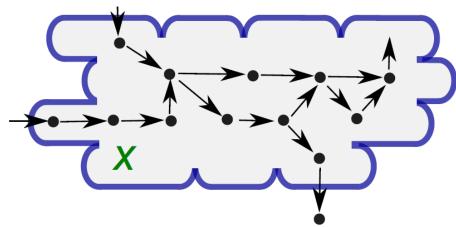


#### **Biochemical reaction networks**

ODE model for growth of microbial populations:

$$\dot{x} = N \cdot v(x) - \mu \cdot x,$$

$$\mu = \delta \cdot \sum_{i} \alpha_{i} \cdot N_{i} \cdot v(x).$$



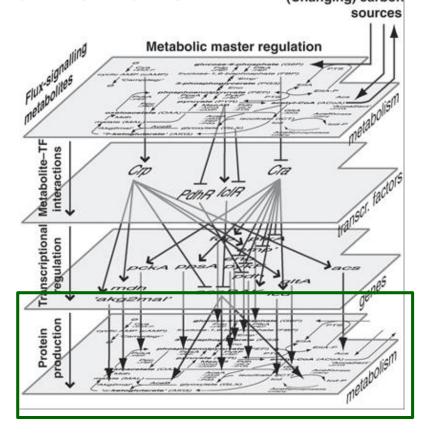
 Reaction rates depend on concentrations x of substrates, products, effectors



#### **Metabolic networks**

- Focus on subsystems that can be studied in isolation due to modular structure of reaction networks

  (Changing) carbon
  - Time-scale hierarchies
  - Connectivity structure
- Metabolic networks
  - Metabolites and enzymatic reactions
  - Short turn-over times of metabolite pools in comparison with enzyme pools



Kotte et al. (2010), Mol. Syst. Biol., 6: 355





#### **Metabolic networks**

- Models describing dynamics of metabolism
  - Effect of growth dilution can often be ignored
  - Variables are metabolites and rates of enzyme-catalyzed reactions
  - Enzyme concentrations constant on time-scale of metabolic dynamics

 $\dot{x} = N v(x)$ 

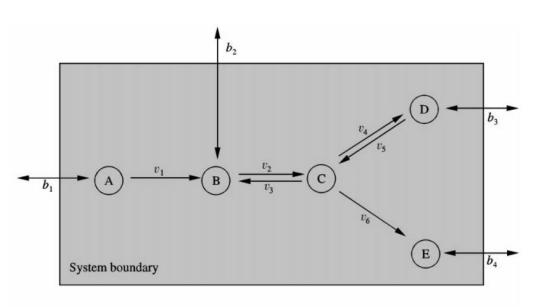


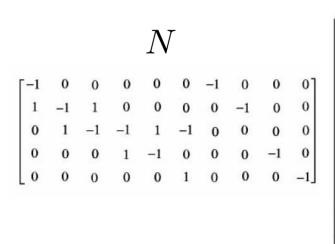


### **Stoichiometry matrix**

• Stoichiometry matrix N describes structure of reaction network

Internal reactions and exchange reactions, reversible and irreversible





Schilling et al. (2000), J. Theor. Biol., 203(3):229-48





Steady state of metabolic network

$$Nv = 0$$

Steady-state reaction rates are called **fluxes** 

Constraints on fluxes: upper and lower bounds

$$v^l \le v \le v^u$$

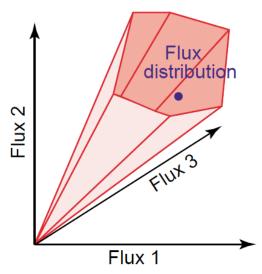
- Bounds on fluxes derived from available information in literature, bounds may be infinite
- For mathematical convenience, all fluxes must be positive  $v \geq 0$
- Reversible reaction modeled as pair of irreversible, positive fluxes



Steady-state dynamics of metabolic network

$$Nv = 0$$

- Stoichiometry matrix and constraints define convex space of possible solutions: flux cone
  - System of steady-state equations underdetermined: more reactions than concentrations variables.
  - Flux cone represents metabolic capabilities of network (possible flux distributions)



Stelling (2004), Curr. Opin. Microbiol., 7:513-8





Steady-state dynamics of metabolic network

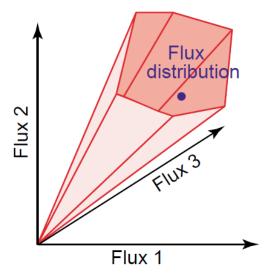
$$Nv = 0$$

- Stoichiometry matrix and constraints define convex space of possible solutions: flux cone
  - System of steady-state equations underdetermined: more reactions than concentrations variables.
  - Every solution can be written as linear combination of rays of flux cone (extreme pathways)

$$C = \{ v \mid v = \sum_{i=1}^{\kappa} w_i p^i, \ w_i \ge 0, \ i = 1, \dots, k \}$$

 $p^i$ : extreme pathway i

 $w_i$ : weigth of *i*th pathway



Stelling (2004), Curr. Opin. Microbiol., 7:513-8





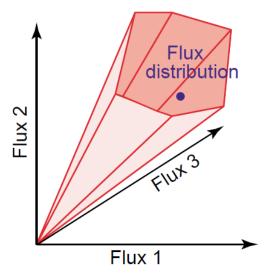
Steady-state dynamics of metabolic network

$$Nv = 0$$

- Stoichiometry matrix and constraints define convex space of possible solutions: flux cone
  - System of steady-state equations underdetermined: more reactions than concentrations variables.
  - Every solution can be written as linear combination of rays of flux cone (extreme pathways)

$$C = \{ v \mid v = \sum_{i=1}^{k} w_i p^i, \ w_i \ge 0, \ i = 1, \dots, k \}$$

 Set of extreme pathways unique, but solutions not uniquely defined by extreme pathways



Stelling (2004), Curr. Opin. Microbiol., 7:513-8



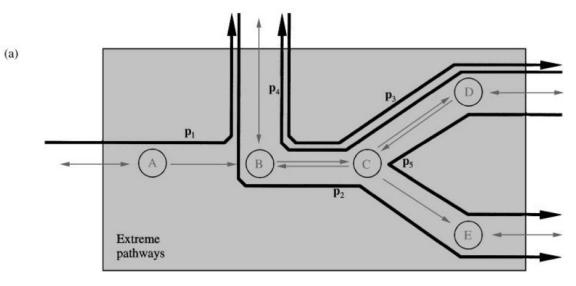


 Extreme pathways in example network

Schilling et al. (2000), J. Theor. Biol., 203(3):229-48

- Extreme pathways provide pathwaybased view of network
- Related concept of elementary modes

Schilling et al. (1999), Biotechnol. Prog., 15(3):296-303



$$\mathbf{P} = \begin{bmatrix} 1 & 0 & 0 & 0 & 0 & 0 & 0 & 0 \\ 0 & 1 & 1 & 0 & 0 & 0 & 0 & 1 \\ 0 & 0 & 0 & 1 & 0 & 0 & 1 \\ 0 & 0 & 1 & 0 & 0 & 1 & 0 \\ 0 & 0 & 0 & 1 & 1 & 1 & 0 \\ 0 & 0 & 0 & 1 & 1 & 1 & 0 \\ 0 & 1 & 0 & 0 & 1 & 0 & 0 \\ \hline -1 & 0 & 0 & 0 & 0 & 0 & 0 \\ 1 & -1 & -1 & 1 & 0 & 0 & 0 \\ 0 & 0 & 1 & -1 & -1 & 0 & 0 \\ 0 & 1 & 0 & 0 & 1 & 0 & 0 \end{bmatrix} \begin{array}{c} v_1 \\ v_2 \\ v_3 \\ v_4 \\ v_5 \\ \hline b_1 \\ b_2 \\ b_3 \\ b_4 \\ \end{array}$$

(c) Flux distribution:  

$$\mathbf{v}^{T} = [4 \ 2 \ 0 \ 1 \ 0 \ 1 - 4 \ 2 \ 1 \ 1]$$

Subset pathways of v:

$${\bf p_1, p_2, p_3}$$

Null space dimensions of Smod:

$$d(S_{mod}) = n - r(S_{mod}) = 8 - 5 = 3$$

Unique decomposition of v:

$$\mathbf{w}^{\mathrm{T}} = [4 \ 1 \ 1 \ 0 \ 0 \ 0 \ 0]$$

$$\mathbf{v} = (4) \cdot \mathbf{p}_1 + (1) \cdot \mathbf{p}_2 + (1) \cdot \mathbf{p}_3$$





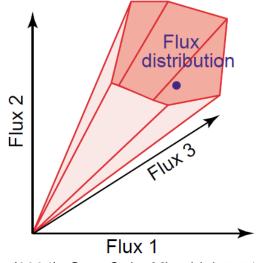
Steady state of metabolic network

$$Nv = 0$$

- Stoichiometry matrix and constraints define convex space of possible solutions: flux cone
- FBA aims at finding solutions maximising or minimising linear combination of fluxes: objective function

$$Z = c^T v \qquad c \in \mathbb{R}^n$$

 Typical objective functions: biomass production, ATP production, ...



Stelling (2004), Curr. Opin. Microbiol., 7:513-8





Steady-state dynamics of metabolic network

$$Nv = 0$$

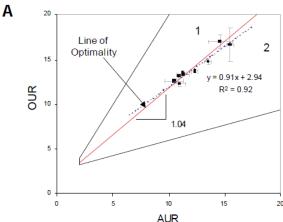
- Stoichiometry matrix and constraints define convex space of possible solutions: flux cone
- FBA aims at finding solutions maximising or minimising linear combination of fluxes: objective function
- Constrained optimisation problem in mathematics
  - Use of LP (linear programming) for solving optimisation problem
  - COBRA toolbox for building and analysing FBA models

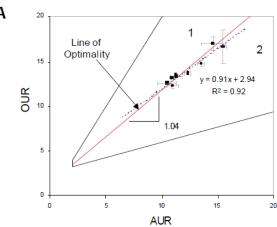
Palsson (2006), Systems Biology: Properties of Reconstructed Networks, Cambridge University Press Orth et al. (2010), Nat. Biotechnol., 28(3):245-8

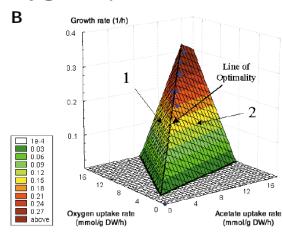




- Genome-scale reconstruction of E. coli metabolism
- FBA predictions of flux distributions maximising growth rate with acetate as carbon source
  - Given acetate and oxygen uptake rates, compute optimal growth rate
  - Line of optimality indicates combinations of acetate and oxygen uptake rates with maximal growth rate
  - Experimental test of predicted line of optimality: control of acetate uptake rate and measurement of growth and oxygen uptake rate







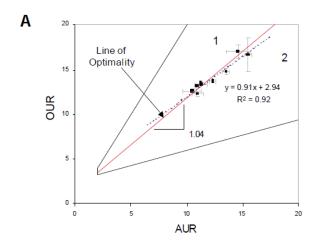




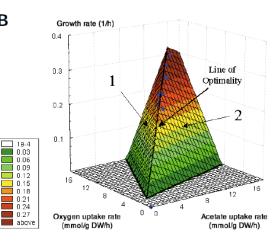
Edwards et al. (2001), Nat. Biotechnol, 19(2):125-30

- Genome-scale reconstruction of E. coli metabolism
- FBA predictions of flux distributions maximising growth rate with acetate as carbon source
- Good correspondence of FBA predictions and experimental data suggests that *E. coli* metabolic network is optimised to maximise growth rate on acetate

Idem succinate





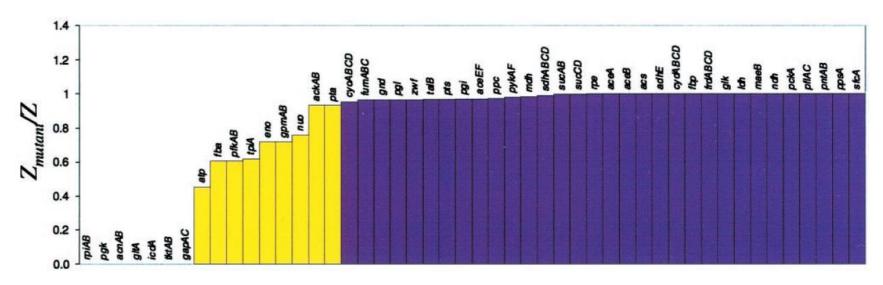


Edwards et al. (2001), Nat. Biotechnol, 19(2):125-30





- Genome-scale reconstruction of E. coli metabolism
- FBA predictions of flux distributions maximising growth rate with glucose as carbon source and fixed oxygen uptake rate
- Effect on growth rate when deleting genes in central carbon metabolism

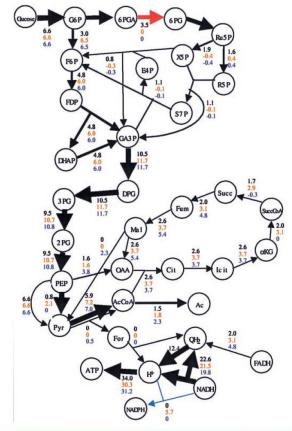


Edwards et al. (2000), Proc. Natl. Acad. Sci. USA, 97(10):5528-33





- Genome-scale reconstruction of E. coli metabolism
- FBA predictions of flux distributions maximising growth rate with glucose as carbon source and fixed oxygen uptake rate
- Effect on flux distribution when deleting genes in central carbon metabolism
  - Deletion of zwf (red) and zwf/pnt (blue)



Edwards *et al.* (2000), *Proc. Natl. Acad. Sci. USA*, 97(10):5528-33





- Genome-scale reconstruction of E. coli metabolism
- FBA predictions of flux distributions maximising growth rate with glucose as carbon source and fixed oxygen uptake rate
- Good correspondence with data for gene deletions examined (86%), but less so for broader range of conditions (60%)
  - Observed growth rate lower than predicted growth rate
- Not surprising: regulatory network of wild-type cells may not be optimal in mutant backgrounds!
  - Regulatory network selects actual flux distribution from possible flux distributions in flux cone





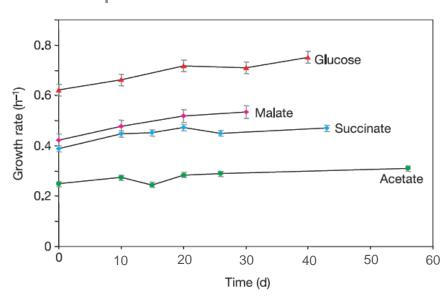
- Genome-scale reconstruction of E. coli metabolism
- FBA predictions of flux distributions maximising growth rate with glucose as carbon source and fixed oxygen uptake rate
- Good correspondence with data for gene deletions examined (86%), but less so for broader range of conditions (60%)
  - Observed growth rate lower than predicted growth rate
- Not surprising: regulatory network of wild-type cells may not be optimal in mutant backgrounds!
  - Regulatory network selects actual flux distribution from possible flux distributions in flux cone
- However, experiments show that E. coli mutant undergoes adaptive evolution to achieve predicted optimal growth rate

Ibarra et al. (2002), Nature, 420(6912):186-9

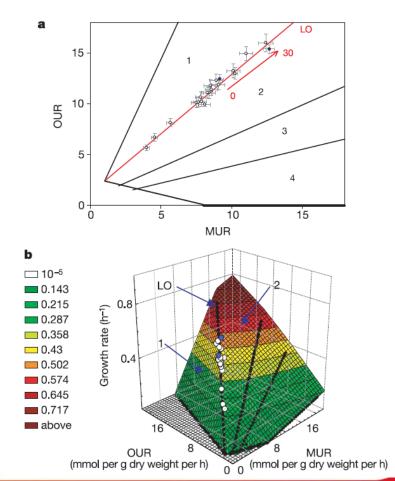




- However, experiments show that E. coli mutant undergoes adaptive evolution to achieve predicted optimal growth rate
  - Growth on malate and other substrates
  - Measured substrate and oxygen uptake rates



Ibarra et al. (2002), Nature, 420(6912):186-9







### Regulatory flux balance analysis

Steady-state dynamics of metabolic network

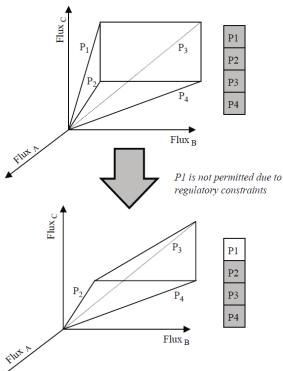
$$Nv = 0$$

• Stoichiometry matrix and constraints define convex space of

possible solutions: flux cone

Refinement of flux cone using additional constraints

Regulation of enzyme activity or expression, switching on/off extreme pathways



Covert et al. (2003), J. Theor. Biol., 221(3):309-25

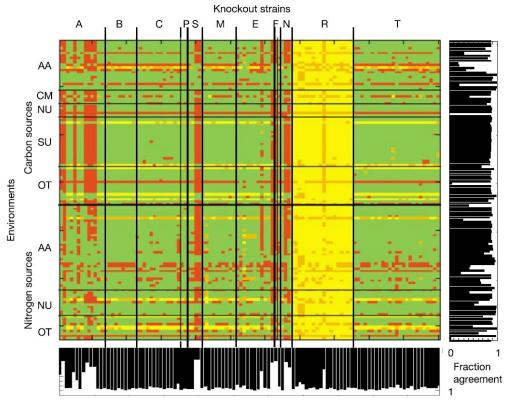




 Regulatory network of wild-type cells may not be optimal in mutant backgrounds

- How do predictions change when including regulatory network?
- Genome-scale model of E. coli metabolism, including regulation of enzymatic genes

Boolean models relating expression of enzymatic genes to growth conditions



Covert et al. (2004), Nature, 429(6987):92-6

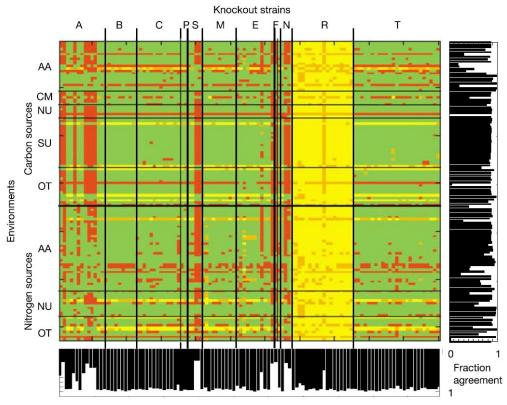




 Regulatory network of wild-type cells may not be optimal in mutant backgrounds

- Genome-scale model of E. coli metabolism, including regulation of enzymatic genes
- Prediction of growth rate in different mutants and growth conditions improved

60% vs 78%



Covert et al. (2004), Nature, 429(6987):92-6





### Dynamic flux balance analysis

Dynamics of metabolic network through interactions with environment

$$\dot{s} = -v_{ext}(t) \cdot B, \quad s(0) = s_0$$
$$\dot{B} = \mu(t) \cdot B, \quad B(0) = B_0$$

B: biomass concentration in medium

s: substrate concentration in medium

 $\mu$ : growth rate

 $v_{ext}$ : substrate uptake rate



- Metabolic network at quasi-steady state with respect to environment
- Computation of exchange rates and growth rate<sub>x</sub>by means of FBA at each time-point t
- Change in substrate concentrations puts bounds on uptake rates

Mahadevan et al. (2002), Biophys. J., 83(3):1331-40





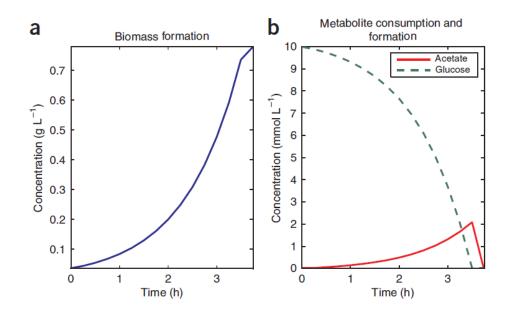
**Biomass** 

# Dynamic flux balance analysis

Dynamics predicted by means of dynamic FBA

Sequential growth of *E. coli* on different carbon sources (glucose, acetate)

Orth et al. (2002), Nat. Protocols, 2(3):727-38







# Monte-Carlo sampling of FBA solutions

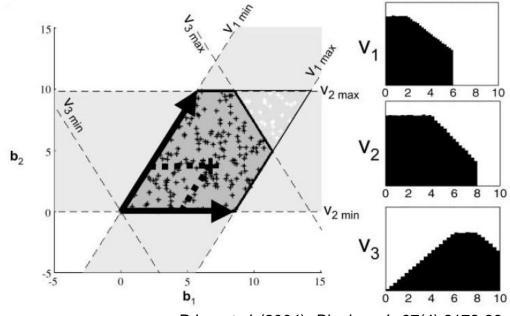
 Stoichiometry matrix and constraints define convex space of possible solutions: flux cone

 FBA selects solutions from flux cone optimizing objective function, but no single solution

Alternative approach: Monte-Carlo sampling of optimal

solutions

Distributions for individual fluxes in network



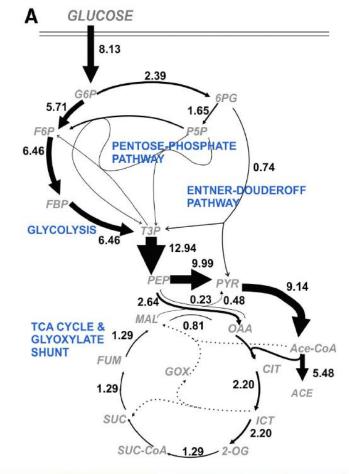
Price et al. (2004), Biophys. J., 87(4):2172-86





### Monte-Carlo sampling of FBA solutions

Analysis of glycolysis pathway in *E. coli* during growth on glucose



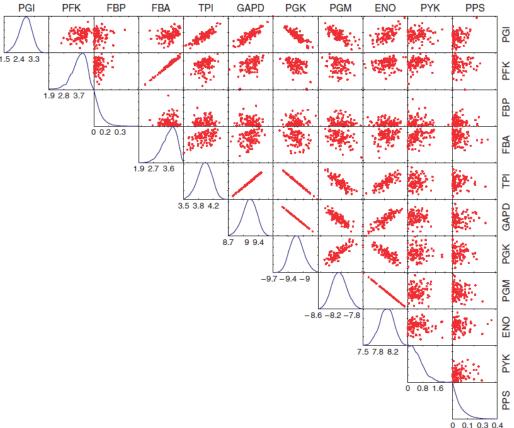


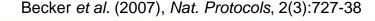


### Monte-Carlo sampling of FBA solutions

Analysis of glycolysis pathway in E. coli during growth on glucose

- Tight distributions
- Correlations between fluxes



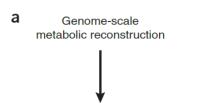


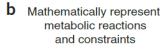




#### **Conclusion FBA**

- FBA models provide genome-scale picture of metabolism and yield experimentally-testable predictions
  - Predictions of flux distributions in different growth conditions and genetic backgrounds







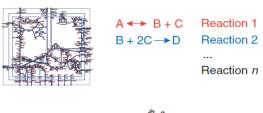
Mass balance defines a system of linear equations

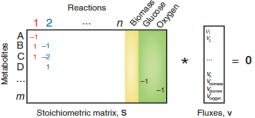


Define objective function  $(Z = c_1^* v_1 + c_2^* v_2 \dots)$ 

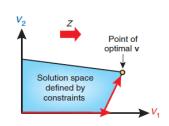


• Calculate fluxes that maximize Z





To predict growth,  $Z = v_{\text{biomass}}$ 



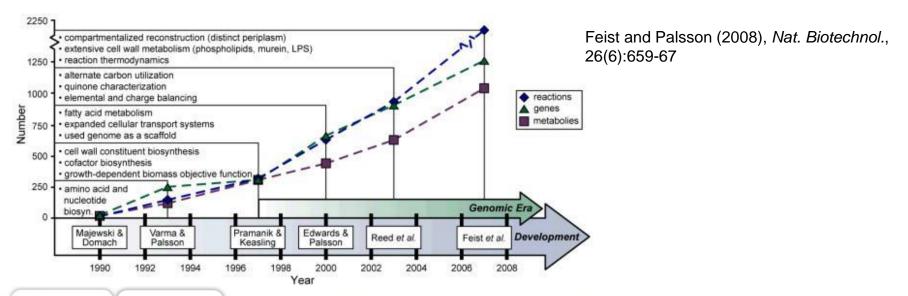
Orth et al. (2010), Nat. Biotechnol., 28(3):245-8





#### **Conclusion FBA**

- FBA models provide genome-scale picture of metabolism and yield experimentally-testable predictions
  - Predictions of flux distributions in different growth conditions and genetic backgrounds
  - Tool for metabolic engineering
  - In E. coli and other (less well-characterised) organisms







#### **Conclusion FBA**

- But FBA has problems as well!
  - Practical question: which objective function works best for problem considered?
  - Fundamental question: what do microorganisms optimise?
     Schuetz et al. (2007), Mol. Syst. Biol., 3:119
  - Integration of regulatory mechanisms on metabolic and genetic level is not easy to achieve in FBA formalism
  - No predictions on dynamics on time-scale of metabolism

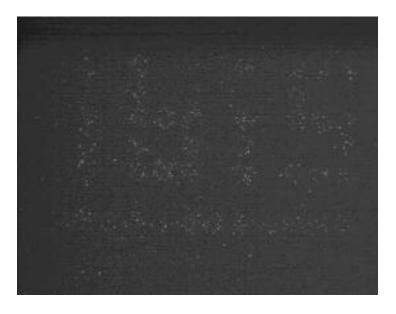




### Internships in IBIS

- Challenging problems for biologists, physicists, computer scientists, mathematicians, ...
- ... in a multidisciplinary working environment
- Contact: Hidde.de-Jong@inria.fr and ibis.inrialpes.fr





Courtesy Guillaume Baptist (2008)





# Merci!



www.inrialpes.fr/ibis